

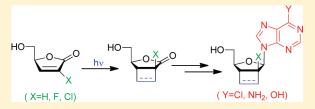
Synthesis of Purine Nucleosides Built on a 3-Oxabicyclo[3.2.0]heptane Scaffold

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Supporting Information

ABSTRACT: The photochemical [2+2] cycloaddition of chiral 3-chloro and 3-fluoro-5-hydroxymethyl-2(5H)-furanone to ethylene and acetylene has been studied. The effect of the halogen atom on the chemical yield and facial diastereoselectivity of the cycloaddition process has been evaluated. From the major anti cycloadducts, practical syntheses of several purine cyclobutane and cyclobutenefused nucleosides containing a halogen atom have been developed. The anti-HIV activity of the new nucleoside analogues has been evaluated.



■ INTRODUCTION

Over several decades, the search for more effective and less toxic antiviral agents against the human immunodeficiency virus (HIV) and hepatitis B virus (HBV) has been focused on 2',3'-dideoxy nucleosides (ddNs). Some successful examples are 3'-azido-2',3'-dideoxythymidine (AZT, zidovudine),¹ 2',3'-dideoxyinosine (ddI, didanosine),² and 2',3'-dideoxycytidine (ddC, zalcitabine) (Figure 1).³ However, side effects, limited stability, and the emergence of drug-resistant mutants remain major drawbacks of these antiviral agents.⁴ Consequently, there is a continuous demand for the identification of structurally new nucleoside derivatives with potent antiviral activity and low toxicity and also for new insights into the structure—activity relationship of these compounds.

The search for new active nucleoside analogues has been primarily focused on modification of the carbohydrate moiety. A structural feature that has often been beneficial for antiviral activity is a 2',3'-unsaturation as in 2',3'-didehydro-2',3'-dideoxythymidine (d4T, stavudine).⁵ Regarding the substituents of the ring, nucleoside analogues with a fluorinated sugar moiety have drawn increasing attention, with many of them showing a broad spectrum of antiviral and anticancer activities.⁶ It is well-known that the introduction of fluorine atoms into an organic molecule often modifies the chemical, physical, and biological properties of the parent compound.⁷

Moreover, it has been found that the introduction of a fluorine atom into the 2'-position of nucleosides results in the stabilization of the glycosyl bond and confers distinctive biological activity. This is a very interesting property, particularly for d2-and d4-purine nucleosides, since it is well established that they are unstable in acidic media, thus limiting their use as orally bioavailable drugs. Accordingly, 2'-fluoro-2',3'-dideoxy nucleosides have been prepared and evaluated as anti-HIV agents and many of them revealed as potent analogues, among which is

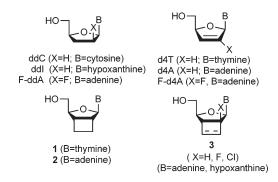


Figure 1. 2',3'-Dideoxynucleosides with recognized activity and novel bicyclic nucleosides built on a 3-oxabicyclo[3.2.0]heptane scaffold.

2'- β -fluoro-2',3'-dideoxyadenosine (F-ddA, Lodenosine). ¹¹ Different 2'-fluoro-2',3'-unsaturated purine analogues have also been synthesized showing potent anti-HIV activity without significant cytotoxicity. ¹²

On the other hand, it is well-known that the conformation and puckering of the glycon moiety of nucleosides play a critical role in modulating their biological activity. Therefore, over the past few years, a considerable amount of work has been done in the synthesis of nucleoside derivatives with fixed sugar-ring puckering, which has provided useful information regarding the relationship between sugar ring conformation and biological activity. In this regard, we have recently described the synthesis and conformational analysis of a novel class of nucleoside analogues, 1 and 2, with the glycone moiety conformationally restricted by a two-carbon chain between positions 2' and 3', which substantially mimic the 2',

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Scheme 1. Synthetic Strategy to Purine Nucleosides Built on a 3-Oxabicyclo [3.2.0] heptane Scaffold

3'-didehydro-2',3'-dideoxynucleoside structure of the well-known antiviral agents d4T and d4A.¹⁵

Considering all these aspects, and in continuation of our efforts to synthesize nucleoside analogues built on a 3-oxabicyclo[3.2.0] heptane scaffold, we turned our attention to the elaboration of the hitherto unknown cyclobutane and cyclobutene purine D-nucleosides of type 3 (Figure 1) containing a heteroatom (such as fluorine or chlorine) at the 2'-position. Herein, we describe the synthesis and anti-HIV activity of such compounds.

■ RESULTS AND DISCUSSION

Our synthetic plan (Scheme 1) involves the diastereoselective construction of the cyclobutane or cyclobutene ring through a [2+2] photochemical reaction of enantiopure 5-hydroxymethyl-2(5*H*)-furanone derivatives 4a-c, the conversion of the cycloadducts 5 to the key intermediates 6, and a subsequent introduction of the selected bases by a Vorbrüggen-type reaction.

Accordingly, our initial efforts focused on the preparation of the bicyclic compounds of type **5**. 2(5*H*)-Furanones **4a**¹⁶ and **4b**¹⁷ were synthesized following procedures described in the literature, while the 3-chloro-2(5*H*)-furanone **4c** was synthesized by a methodology similar to that used to prepare its fluoro analogue (Scheme 2). Thus, the Horner–Emmons reaction of 2,3-*O*-isopropylidene-D-glyceraldehyde, 7, with triethyl 2-chloro-2-phosphonoacetate and BuLi in dry THF afforded a chromatographically separable mixture of olefins **8** and **9** (86:14) in 62% yield. The use of other bases such as sodium hexamethyldisilyamide (NaHMDS) proved to be less effective for this transformation. Acid treatment of the major stereoisomer **8** delivered **4c** in 95% yield.

The cyclobutane derivative 11 has been efficiently prepared by our two-step protocol le consisting of the photochemical [2+2] cycloaddition of lactone 4a with (Z)-1,2-dichloroethylene to afford a mixture of dichlorocycloadducts 10, followed by a dihydrodehalogenation reaction of 10 by treatment with n-tributyltin hydride and AIBN in refluxing toluene. This protocol delivered a 90:10 mixture of the cyclobutanes 11 and 12 in 73% overall yield (Scheme 3). Alternatively, treatment of the mixture of 10 with activated Zn in 80% EtOH in a sealed vessel at 100 °C, under microwave irradiation (microwave reactor, IR temperature monitoring), furnished a mixture of the cyclobutene diastereomers 13 and 14 in 90:10 ratio and 75% yield, wherefrom the major isomer 13 could be isolated by column chromatography in 65% yield. The structure of the products was assigned by 1D and 2D NMR spectroscopy. The value of the coupling constant

Scheme 2. Synthesis of the Chlorolactone 4c

Scheme 3. Preparation of 11 and 13

between H-4 and H-5 is diagnostic for the anti/syn relative configuration of the cycloadducts. A small value of $J_{4,5}$ (1.7 Hz) is in agreement with a trans relationship between these two protons, whereas a larger value (4.3 Hz) indicates a cis relationship.¹⁹

Next, we turned our attention to the synthesis of the 3-oxabicyclo[3.2.0]heptane scaffold bearing a fluoride or chloride atom at the α -carbonyl position in which the presence of the halogen atom precludes the utilization of the above protocol. Therefore, we decided to prepare the halo derivatives by photochemical [2 + 2] cycloaddition of the corresponding 3-halo-2(5H)-furanones 4b and $4c^{20}$ to ethylene and acetylene. So far, there have been limited reports on the photochemical behavior of α , β -unsaturated ketones or lactones containing a vinylic halogen atom, and to the best of our knowledge, photochemical reactions of 3-halo-2(5H)-furanones with unsaturated substrates have not been reported yet. The influence of the α -halogen atom over the reactivity and stereoselectivity has been evaluated using as a control the photoreaction of lactone 4a with ethylene and acetylene (Scheme 4). The results are summarized in Table 1.

First, we studied the photoreaction of lactones 4a-c with ethylene. The photoreaction of lactone 4b (entry 2) afforded an 81:19 mixture of the anti:syn isomers 15 and 16 from which the major isomer 15 was isolated in 62% yield. Thus, the presence of a vinylic fluorine atom enhanced both the yield and the facial selectivity compared to the results with lactone 4a (entry 1). In contrast, the photochemical reaction of chlorolactone 4c (entry 3) furnished a 72:11:17 mixture of three isomers in 85% global yield. Purification by column chromatography allowed the separation of all three products, which were identified as the anti cycloadduct 17 (58%) and the two spirocyclopropane lactones 23 (8%) and 24 (13%) (Figure 2). No presence of the syn cycloadduct 18 was observed in the crude material.

Scheme 4. [2+2] Photochemical Cycloaddition of 4a-c to Ethylene and Acetylene

The structure determination of the cyclopropane derivatives 23 and 24 was accomplished on the basis of their ¹H and ¹³C NMR spectra with the help of 2D NMR experiments. For both compounds, the COSY experiments revealed two independent sets of signals, one of them containing four cyclopropane protons between 1.3 and 1.6 ppm $(2 \times H-1)$ and $(2 \times H-2)$ and the second containing the remaining four protons between 3.8 and 4.8 ppm (H-6, H-7, and $2 \times$ H-8). The HMQC experiments showed the presence of a quaternary carbon atom at 28.6 ppm for 23 and 28.1 ppm for 24, which were assigned to the spiranic position C-3. On the other hand, the HMBC experiments showed correlations between H-7 and C-1, C-2, C-3, and C-6 and between the cyclopropane protons H-1/H-2 and C-4. Finally, the configuration of the new stereogenic center C-7 was assigned considering the value of the coupling constant between H-6 and H-7, which is 5.0 Hz for 23, indicating a cis disposition between H-6 and H-7, and 3.3 Hz for 24, denoting a trans relationship.

The formation of the spirocyclopropyl derivatives 23 and 24 might be rationalized considering the postulated isomeric 1,4-biradical intermediates I—IV involved in the photochemical reaction (Scheme 5). Intermediates I and II would deliver the anti cycloadduct 17, while III and IV direct to the syn isomer 18. It has been suggested that the product distribution is controlled by the relative amount of the different isomeric 1,4-biradicals formed as well as by the extent to which each of them partitions between closure to the product and reversion to the precursors. ^{22,23} In this case, however, the 1,4-biradical intermediates II and III may evolve through a competitive pathway involving an intramolecular chlorine migration leading to the rearranged 1,3-biradical intermediates II' and III' that will eventually furnish the spirocyclopropyl lactones 23 and 24. It is noteworthy to mention that, in this reaction, the formation of the syn cycloadduct 18 is not

HO
$$\frac{8}{6}$$
 $\frac{0}{3}$ $\frac{4}{2}$ $\frac{0}{2}$ HO $\frac{0}{2}$ $\frac{0}{2}$ $\frac{0}{24}$

Figure 2. Spirocyclopropane lactones 23 and 24.

Scheme 5. Formation of the Spirocyclopropyl Lactones 23 and 24

observed, meaning that the intermediate radical III leads exclusively to the spiro lactone **24.** Likely, due to the trans disposition of the hydroxymethyl chain and the chlorine atom in III, the intramolecular abstraction of the vicinal chlorine atom is favored over the cyclization process when compared with the analogous radical II, where a more sterically demanding situation may disfavor the chlorine migration.

Next, the [2+2] photocycloaddition of 4a-c to acetylene was targeted to our attention (Table 1, entries 4-6). These photoreactions proceeded in much lower chemical yields. Thus, irradiation of an acetone solution of 4a and acetylene for 5.3 h furnished a 65:35 mixture of the cyclobutene adducts 13 and 14 in 53% total yield (entry 4). The photoreaction of the fluorolactone 4b (entry 5) afforded a complex crude material from which a 95:5 mixture of adducts 19 and 20 was isolated (28%), along with the unexpected tricyclic lactone 25 (15%, Figure 3). Likewise, the chlorolactone 4c furnished the tricyclic lactone 26 (6%), besides the anti cycloadduct 21 (17%) (entry 6). In this case, the primary syn cycloadduct 22 was not even observed.

Table 1. [2+2] Photocycloaddition of 2(5H)-Furanones 4a-c to Ethylene and Acetylene

entry	lactone	X	unsaturated substrate	t	cycloadduct yield b (%)	anti:syn ^c (%)	other products d
1	4a	Н	ethylene	5.5 h	66	66:34	
2	4b	F	ethylene	3.5 h	80	81:19	
3	4c	Cl	ethylene	4.5 h	58	100:-	23 (8%), 24 (13%)
4	4a	Н	acetylene	5.3 h	53	65:35	
5	4b	F	acetylene	5.5 h	28	95:5	25 (15%)
6	4c	Cl	acetylene	4.5 h	17	100:-	26 (6%)

^a Substrates 4a-c in acetone solutions saturated with ethylene or acetylene were irradiated through a Pyrex filter with a 125 W high-pressure mercury lamp at -20 °C. ^b Isolated yield of the mixture of stereoisomers after column chromatography. ^c Product ratio determined by GC and ¹H NMR. ^d Isolated yield after column chromatography.

Figure 3. Tricyclic photoproducts.

Scheme 6. Preparation of the Cyclobutene Adenine Analogues

The lower yields of the photochemical reactions with acetylene compared to the corresponding reactions with ethylene can be attributed to secondary photoactivity since the initial cyclobutene photoproducts might undergo excitation leading to further processes, which can also mask the observed anti/syn selectivity of the photocycloaddition. ²⁴ The tricyclic cyclobutene products **25** and **26** might arise from the reaction of the newly formed double bond of the primary [2+2] adducts with acetylene. Formation of tricyclic cyclobutene derivatives through two consecutive additions of acetylene had been already reported by Pietra and co-workers ²⁵ in the photochemical reaction of 3-acetoxy-2-cyclopentenone with acetylene.

The structural and stereochemical assignments of the cycload-ducts 25 and 26 were established on the basis of 1D and 2D NMR spectroscopy. For instance, for compound 25, the HMQC experiment showed two allylic methyne groups, C-2 and C-5, while the HMBC spectrum showed correlations between H-1 and C-2, C-3, C-6, and C-10, between H-2 and C-3, C-4, and C-5, between H-4 and C-3 and C-5, and between H-5 and C-6. The relative configuration was determined on the basis of NOESY experiments. Thus, the cross-peak observed between H-3 and H-9 indicates an anti endo stereochemistry.

In summary, the presence of a halogen atom at the α -position of the lactone has a noticeable influence on the chemical yield and stereochemical course on their photochemical reactions with ethylene and acetylene. Considering all the factors implicated in the studied photochemical processes, only the photoreactions involving ethylene are synthetically useful. These reactions furnished the fluoro- and chlorocyclobutanes 15 and 17 in 62% and 58% yield, respectively.

With the major anti cycloadducts 13, 15, and 17 in hand, we turned our attention to prepare the target nucleoside analogues. First, we undertook the synthesis of the purine cyclobutene analogues starting from 13 (Scheme 6).

Scheme 7. Preparation of the Fluoro and Chloro Cyclobutane Purine Analogues

According to the plan, the protection of the primary hydroxyl group as the *tert*-butyldimethylsilyl ether, followed by partial reduction of the lactone with DIBAL-H in toluene, and treatment of the resulting lactols with acetic anhydride provided the key intermediate 27 in 64% yield for the three steps.

The N-glycosylation reaction of 27 with 6-chloropurine was conducted under modified Vorbrüggen conditions; ²⁶ namely, 6-chloropurine was silylated in situ by reaction with N,O-bis-(trimethylsilyl)acetamide (BSA) in the presence of trimethylsilyltriflate (TMSOTf) in CH₃CN at room temperature. Under these conditions, the condensation reaction proceeded with the formation of the expected N9 α - and β -anomers 28 and 29 in 86% overall yield and nearly equal amounts. The chromatographic separation of 28 and 29 provided pure samples of the two epimers. Since the α -anomer might also be a valuable candidate for biological assays, alternative conditions to increase the β -selectivity of the coupling reaction were not investigated.

The *N*9 attachment site of the purine base in **28** and **29** was deduced from HMBC experiments, which showed correlation between H-1' and C-4, while the anomeric configuration was elucidated by ¹H NMR analysis, including NOESY experiments.²⁷ The isomer displaying a small coupling constant $J_{1',2'}$ (\sim 0 Hz) was assigned as the β -anomer **29**, whereas that with a larger $J_{1',2'}$ (5.4 Hz) was assigned as the α-anomer **28**.

Unmasking of the hydroxyl group of **29** to furnish **31** was accomplished by routine desilylation with tetra-*n*-butylammonium fluoride. Finally, substitutive amination by heating **31** at 90 $^{\circ}$ C with methanolic ammonia in a sealed vessel for 8 h delivered the targeted cyclobutene nucleoside **33** in 72% yield. An analogous sequence from **28** provided the α -anomer **32** in 72% yield for the two steps.

The synthesis of the fluoro and chloro cyclobutane analogues was carried out in a similar manner (Scheme 7). Thus, the anti cycloadducts 15 and 17 were converted to the corresponding fluoro acetates 34–35 and chloro acetate 36 in 78% and 82% overall yield, respectively.

Table 2. Different Reaction Conditions for the N-Glycosylation Reaction

entry	X	solvent	T	time	yield (%) ^a	$N9\alpha:N9\beta:N7\alpha:N7\beta$ (%) ^b	N-9:N-7	α:β
1	Cl	acetonitrile	25 °C	3.5 h	87	42:4:47:7	1:1.2	8.1:1
2	Cl	acetonitrile	82 °C	1 h	95	74:13:8:5	6.7:1	4.5:1
3	Cl	DCE	25 °C	27 h	65	31:1:58:10	1:2.1	8.1:1
4	Cl	DCE	83 °C	0.5 h	78	70:11:13:6	4.3:1	4.9:1
5	Cl	toluene	25 °C	16 h	38	39:5:50:6	1:1.3	8.1:1
6	Cl	toluene	100 °C	1 h	77	85:15:-:-	1:0	5.6:1
7	F	acetonitrile	25 °C	7.5 h	88	28:15:35:22	1:1.3	3.7:1
8	F	DCE	83 °C	2.5 h	77	34:20:23:23	1.2:1	1.3:1
9	F	toluene	100 °C	1 h	90	59:35:3:3	15:1	1.6:1
^a Isolated yield after column chromatography. ^b Product ratio was determined by ¹ H NMR.								

At first, the N-glycosylation reaction of acetates 34-35 and 36 with silylated 6-chloropurine was attempted in acetonitrile at rt, under the same coupling conditions previously used for 27 (Table 2, entries 1 and 7). Surprisingly, in these cases, the condensation reaction resulted in the formation of the expected N9 α - and β -isomers (37 and 38 for X = F and 41 and 42 for X = Cl) along with the corresponding N7 coupling products (39 and 40 for X = F and 43 and 44 for X = Cl). All the isomers were isolated by column chromatography, and their regio- and stereochemistry were assigned by 1D and 2D NMR spectroscopy.

The attachment site of the purine base in the N9 isomers $37{-}38$ and $41{-}42$ was established by HMBC experiments, which showed correlation between H-1' and the aromatic carbon atoms C-4 and C-8. On the other hand, cross-peaks were observed between H-1' and C-5 and C-8 for 40, indicating N7 connectivity. However, no cross-peak was observed between H-1' and C-5 for compounds 39, 43, and 44. In these compounds, the N7 connectivity was supported by the higher chemical shift value of the purine C-4 carbon of the N7 isomers $39{-}40$ and $43{-}44$ (~ 162 ppm) compared to that of the N9 isomers $37{-}38$ and $41{-}42$ (~ 151 ppm), due to the larger steric congestion existing in the latter.

The anomeric configuration of the fluoro derivatives 37-40 was established through the analysis of their 1H and ^{13}C NMR spectra. The value of $^2J_{F,C-1'}$ is a useful tool to determine the configuration of the anomeric position of the 2'-fluoronucleosides. 29 Thus, the $^2J_{F,C-1'}$ coupling constant values for β -isomers are close to 18 Hz, whereas for the α -isomers they are around 40 Hz. Accordingly, the highest values of the carbon—fluorine coupling constant between C-1' and F were correlated with the α -anomeric configuration (40.7 Hz for 37 and 38.7 Hz for 39); meanwhile, the lowest values were correlated with the β -anomeric configuration (17.8 Hz for 38 and 18.1 Hz for 40). Moreover, both N9 and N7 β -nucleosides 38 and 40 possess a characteristic long-range coupling between the fluorine atom and the

H-8 proton of the purine base that is not observed in their corresponding $\alpha\text{-epimers.}^{30}$

The anomeric configuration of the chloro derivatives 41-44 was established by NOESY experiments. Thus, the α -anomers 41 and 43 showed clear correlations between H-1' and H-5', while the β -anomers 42 and 44 showed correlations between H-1' and H-4'.

In view of these results and to overcome the undesired N7 regioselectivity, alternative coupling reaction conditions were investigated. The effect of the solvent 26 and the temperature 31 on the outcome of the reaction was evaluated (Table 2). Eventually, it was found that by performing the N-glycosylation reaction in toluene at 100 °C the N9 isomers were obtained as major products in good yields (entries 6 and 9). Thus, under these conditions, the fluoro acetates 34-35 delivered a 15:1 mixture of the α/β N9 isomers 37 and 38 and the α/β N7 isomers 39 and 40 in 90% overall yield, while the reaction of the chloro acetate 36 proceeded with complete regioselectivity affording a mixture of the α/β N9 isomers 41 and 42 in 77% overall yield. With regard to the α/β stereoselectivity of the condensation reaction, the fluoro derivative gave an $\alpha:\beta$ ratio of 1.6:1, whereas that for the chloro derivative was 5.6:1, suggesting that the chlorine atom acts as an important steric factor in the coupling reaction.

Next, removal of the silyl protecting group of the α -anomers 37 and 41 by treatment with Et₃N·3HF in THF furnished the alcohols 45 and 47 in 90% and 88% yield, respectively (Scheme 7). Ammonolysis of these intermediates gave the α -adenosine analogues 49 and 51 in 89% and 91% yield, respectively. The same protocol was applied to the β -anomers 38 and 42 to afford the corresponding target β -adenosine analogues 50 and 52 in 68% and 87% yield for the two steps, respectively.

For these compounds, the substitution of the aromatic chlorine atom of the fluoro, 45-46, and the chloro derivatives, 47-48, by a hydroxyl group was also attempted. One of the most

Table 3. Some Conformational Parameters of 33, 2, ddA, d4A, and 53

compd	P^a (deg)	$v_{\mathrm{max}}^{}b}\left(^{\circ}\right)$	χ^{c} (°)	γ^d (°)	d^e (Å)
33	259.8	25.7	-175.5	-75.6	3.4 (4.8)
2	228.1	22.9	-157.1	173.8	3.7 (4.0)
ddA	190.4	35.7	-95.9	-178.7	3.9 (4.5)
d4A	243.5	7.5	-100.2	179.8	3.9 (4.6)
53	247.1	27.5	163.6	56.6	

 aP : phase angle of pseudorotation. $^bv_{\rm max}$: maximum puckering amplitude. $^c\chi$: torsion angle O4′–C1′–N9–C4. $^d\gamma$: torsion angle O5′–C5′–C4′–C3′. ed : distance between C-5′ and N9 (distance between O5′ and N9).

common methodologies to achieve this transformation is based on the treatment of a 6-chloropurine derivative with 2-mercaptoethanol and sodium methoxide in refluxing aqueous methanol. However, all the attempts to prepare the hypoxanthine derivatives using these conditions met with failure. It was eventually found that performing the reaction in refluxing dioxane the targeted fluoro and chloro inosine analogues 53–54 and 55–56 could be obtained in good yields.

The adenine cyclobutene β -anomer 33 provided adequate crystals for X-ray analysis. Unfortunately, we were unable to grow good-quality crystals for the fluoro and chloro cyclobutane β -anomers, and within this series, only the 2'-fluoro α -hypoxanthine analogue 53 furnished suitable crystals for X-ray analysis. From the crystallographic data of 33 and 53, we have calculated their pseudorotational parameters, which revealed that the furanose ring of the cyclobutene analogue 33 adopts an O-exo ($_{O}E$) pucker with a pseudorotational phase angle $P=259.8^{\circ}$ and a maximum amplitude of puckering $v_{\rm max}=25.7^{\circ}$, while the furanose ring of 53 presents a C-4' endo (^{4}E) pucker with a pseudorotation phase angle $P=247.1^{\circ}$ and a puckering amplitude of $v_{\rm max}=27.5^{\circ}$.

The pseudorotational parameters of 33 have been compared to those obtained from the crystallographic data of 2,15 d4A,37 and ddA³⁸ (Table 3). A close inspection of these parameters showed several significant differences, but all four compounds are settled in the southwest region of the pseudorotational cycle. In terms of puckering, the furanosid ring of 33 ($v_{\rm max}$ = 25.7°) is not as planar as that of d4A ($v_{\rm max}$ = 7.5°), but it is flattened compared to ddA ($v_{\rm max}$ = 35.7°), indicating that the cyclobutene motif limits the conformational flexibility of the sugar moiety. Moreover, the puckering of the furanoside ring of 33 is similar to that of 2 ($v_{\text{max}} = 22.9^{\circ}$), showing that the pursued additional rigidity induced by the double bond in 33 is not at play. However, puckering amplitude values around 25° are close to a "central conformation" which includes nucleosides with small $v_{\rm max}$ values of less than 20°. With regard to the glycosidic link, although the purine base in all compounds is anti arranged relative to the sugar moiety, the glycosyl torsion angle χ values of 33 (-175.5°) and 2 (-157.1°) clearly diverge from those of d4A (-100.2°) and ddA (-95.9°) . The main difference between 33 and 2 is the orientation of the hydroxymethyl group (Figure 4). Thus, the value of the torsion angle γ of 33 (-75.6°) is strikingly different from those of 2, d4A, and ddA (\sim 178°). Finally, another remarkable conformational feature of 33 is that the distance between C5' and N9, an important parameter for the recognition by nucleoside kinases, is comparable to those of 2, ddA, and d4A.³⁹

As a preliminary test, the new synthesized nucleoside analogues were evaluated on MT4 cells for anti-HIV-1 activity against

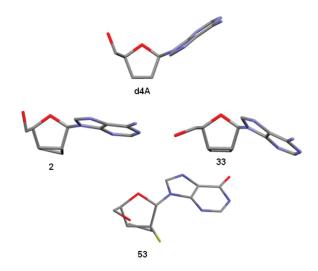


Figure 4. X-ray structures of d4A, 2, 33, and 53.

wild-type NL4-3 strain as well as cytotoxicity using d4T and AZT and AMD3100 as the positive control. Unfortunately, only a moderate activity was found for 2 (EC₅₀ = 17.8 μ g/mL and CC₅₀ > 25 μ g/mL) and 52 (EC₅₀ = 22.92 μ g/mL and CC₅₀ > 25 μ g/mL), and no activity or toxicity was found for any of the remaining compounds below a concentration of 25 μ g/mL.⁴⁰ Despite the similar structural motifs of the synthesized compounds and d4A, the weak anti-HIV-1 activity displayed by our compounds may suggest that the ethylene group of the fused cyclobutane makes them poor substrates for the three activating cellular kinases or for the final interaction with the target reverse transcriptase.

CONCLUSIONS

In summary, purine nucleoside analogues 32-33 and 49-56 built on a 3-oxabicyclo[3.2.0]heptane scaffold bearing different functionality have been synthesized through glycosylation reactions between 6-chloropurine and suitable sugar precursors containing the 3-oxabicyclo[3.2.0]heptane skeleton 13, 15, and 17. The anti cyclobutene derivative 13 has been stereoselectively prepared by the photoreaction of 4a with (Z)-1,2-dichloroethylene followed by Zn-promoted dechlorination under microwave irradiation, while the halogenated anti cyclobutanes 15 and 17 have been efficiently prepared by the photochemical reaction of lactones 4b and 4c with ethylene. The presence of a halogen atom at the α -position of the lactone has a noticeable influence on the outcome of the photochemical process. Structure, conformation, and anti-HIV-1 activity of the new nucleosides have been investigated.

■ EXPERIMENTAL SECTION

General Methods. Commercially available reagents were used as received. The solvents were dried by distillation over the appropriate drying agents. All reactions were performed avoiding moisture by standard procedures and under a nitrogen atmosphere. Flash column chromatography was performed using silica gel (230–400 mesh). 1 H NMR and 13 C NMR spectra were recorded at 250 and 62.5 MHz or 360 and 90 MHz or 500 and 125 MHz. NMR signals were assigned with the help of DEPT, COSY, HMBC, HMQC, and NOESY experiments. Proton chemical shifts are reported in parts per million (δ) (CDCl₃, δ 7.26, CD₃OD δ 3.31, or DMSO- d_{6} , δ 2.50). Carbon chemical shifts are

reported in parts per million (δ) (CDCl₃, δ 77.2, CD₃OD δ 49.00, or DMSO- d_6 , δ 39.52). NMR signals were assigned with the help of COSY, HSQC, HMBC, and NOESY experiments. Infrared peaks are reported in cm⁻¹. Melting points were determined on hot stage and are uncorrected. High-resolution mass spectra (HRMS) were recorded on a mass spectrometer. Optical rotations were measured at 22 \pm 2 °C.

Microwave reactions were conducted on a microwave synthesizer. The machine consists of a continuous focused microwave-power delivery system with operator-selectable power output from 0 to 300 W. The temperature of the contents of the vessel was monitored using a calibrated infrared temperature control mounted under the reaction vessel. All experiments were performed using a stirring option whereby the contents of the vessel were stirred by means of a rotating magnetic plate located below the floor of the microwave cavity and a Teflon-coated magnetic stir bar in the vessel.

Anti HIV-1 Activity and Cytotoxicity. Anti-HIV activity and citotoxicity were assayed at the Irsicaixa Foundation, University Hospital Germans Trias i Pujol, Badalona, Spain. Anti-HIV activity was tested in a lymphoid cell line MT-4 which was obtained through the MRC AIDS Reagent Program. Briefly, the HIV-1 NL4-3 strain was tittered in MT-4 cells after acute infection and infectivity were measured by evaluating the cytopathic effect induced after 5 day cultures as described. Anti-HIV activity and cytotoxicity measurements in MT-4 cells were based on the viability of cells that had been infected or not infected with HIV-1, all exposed to various concentrations of the test compound. After the MT-4 cells were allowed to proliferate for 5 days, the number of viable cells was quantified by a tetrazolium-based colorimetric method (MTT method) as described.

Ethyl (2E)-3-[(4S)-2',2'-Dimethyl-1',3'-dioxolan-4'-yl]-2-chloro-2propenoate (8) and Its (2Z)- Isomer (9). To a -78 °C cooled solution of triethyl 2-chloro-2-phosphonacetate (1.02 g, 3.93 mmol) in dry THF (2 mL) was added dropwise BuLi (2.44 mL, 3.91 mmol, 1.6 M) in hexane, and the solution was stirred for 1 h at -78 °C. Then, a solution of 2,3-O-isopropylidene-D-glyceraldehyde, 7 (510 mg, 3.91 mmol), in dry THF (2 mL) was added dropwise, and the dark solution was kept at -78 °C for 2 h. The reaction mixture was treated with a saturated aqueous solution of NH₄Cl (2 mL) and was allowed to slowly warm to room temperature. After addition of CH₂Cl₂ (5 mL), the aqueous layer was extracted with CH_2Cl_2 (3 × 2 mL). The combined organic layer was washed with brine (3 × 4 mL), dried over anhydrous Na₂SO₄, and evaporated. The resulting dark oil was purified by column chromatography (from hexane to hexane—diethyl ether 30:1) affording the olefins 8 (489 mg, 2.08 mmol, 53% yield) and 9 (80 mg, 0.34 mmol, 9% yield) as colorless oils.

8: IR (ATR) 2985, 2937, 1714, 1208, 1058, 1024 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 6.57 (d, $J_{3,4'}$ = 6.9 Hz, 1H, H-3), 5.29 (ddd, $J_{4',3}$ = 6.9 Hz, $J_{4',5'}$ = 6.8 Hz, $J_{4',5'}$ = 6.3 Hz, 1H, H-4'), 4.32 (dd, J_{gem} = 8.4 Hz, $J_{5',4'}$ = 6.8 Hz, 1H, H-5'), 4.28 (m, 2H, OCH₂CH₃), 3.70 (dd, J_{gem} = 8.4 Hz, $J_{5',4'}$ = 6.3 Hz, 1H, H-5'), 1.45 (s, 3H, CH₃), 1.38 (s, 3H, CH₃), 1.35 (t, J = 7.1 Hz, 3H, OCH₂CH₃); ¹³C NMR (90 MHz, CDCl₃) δ 162.0 (C=O, C-1), 144.6 (CH, C-3), 123.7 (C, C-2), 110.0 (C, C-2'), 73.6 (CH, C-4'), 69.3 (CH₂, C-5'), 62.5 (CH₂, OCH₂CH₃), 26.5 (CH₃, C(CH₃)₂), 25.4 (CH₃, C(CH₃)₂), 14.0 (CH₃, OCH₂CH₃). HRMS (ESI+) calcd for [C₁₀H₁₅ClO₄ + Na]⁺ 257.0551, found 257.0547 (mixture of 8 and 9).

9: IR (ATR) 2985, 2929, 1726, 1240, 1060, 1030 cm $^{-1}$; 1 H NMR (360 MHz, CDCl₃) δ 7.05 (d, $J_{3,4'}$ = 6.9 Hz, 1H, H-3), 4.97 (ddd, $J_{4',3}$ = 6.9 Hz, $J_{4',5'}$ = 6.8 Hz, $J_{4',5'}$ = 6.8 Hz, 1H, H-4), 4.24 (m, 3H, H-5', OCH₂CH₃), 3.66 (dd, J_{gem} = 8.4 Hz, $J_{5',4'}$ = 6.8 Hz, 1H, H-5'), 1.40 (s, 3H, CH₃), 1.35 (s, 3H, CH₃), 1.29 (t, J = 7.1 Hz, 3H, OCH₂CH₃); 13 C NMR (90 MHz, CDCl₃) δ 161.5 (C=O, C-1), 140.0 (CH, C-3), 125.7 (C, C-2), 110.1 (C, C-2'), 73.3 (CH, C-4'), 68.1 (CH₂, C-5'), 62.5 (CH₂, OCH₂CH₃), 26.3 (CH₃, C(CH₃)₂), 25.4 (CH₃, C(CH₃)₂), 14.0 (CH₃, OCH₂CH₃).

(55)-3-Chloro-5-hydroxymethyl-2(5H)-furanone (**4c**). To a solution of **8** (300 mg, 1.28 mmol) in EtOH (10 mL) at 0 °C was added concentrated HCl (0.4 mL, 5.12 mmol). The mixture was allowed to warm to room temperature and stirred for 4 h. Then, the solvent was removed and the residue coevaporated with toluene (2 × 2 mL). The resulting crude was purified by column chromatography (EtOAc) yielding **4c** (180 mg, 1.21 mmol, 95% yield) as a colorless oil: [α]_D – 54.0 (c 1.0, CHCl₃); IR (ATR) 3409, 3097, 1752, 1029 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 7.35 (d, $J_{4,5}$ = 1.9 Hz, 1H, H-4), 5.12 (ddd, $J_{5,6}$ = 4.8 Hz, $J_{5,6}$ = 3.8 Hz, $J_{5,4}$ = 1.9 Hz, 1H, H-5), 4.00 (dd, J_{gem} = 12.3 Hz, $J_{6,5}$ = 3.8 Hz, 1H, H-6), 3.83 (dd, J_{gem} = 12.3 Hz, $J_{6,5}$ = 4.8 Hz, 1H, H-6), 2.37 (br s, 1H, OH); ¹³C NMR (62.5 MHz, CDCl₃) δ 167.7 (C=O, C-2), 145.0 (CH, C-4), 126.1 (C, C-3), 81.5 (CH, C-4), 62.3 (CH₂, C-6). HRMS (ESI+) calcd for [C₅H₅ClO₃ + Na]⁺ 170.9819, found 170.9825.

General Procedure for the Photochemical Reactions. Irradiations were performed in a small conventional photochemical reactor (two-necked vessel fitted with a Pyrex or quartz immersion type cooling jacket) using a high-pressure 125 W mercury lamp. Methanol at $-15\,^{\circ}$ C was used for refrigeration of the immersion well jacket. The vessel was externally cooled at $-20\,^{\circ}$ C with a dry ice—CCl₄ bath or at $-40\,^{\circ}$ C with a dry ice—acetonitrile bath. Ethylene or acetylene (acetone free) gas was bubbled through the solution for 15 min. Once the lamp was turned on, a slow flow of gas was maintained throughout the irradiation. *Caution: acetylene is hazardous and subject to strict safety regulations.* The photochemical reactions with (Z)-1,2-dichloroethylene were initially degassed by passage of oxygen-free argon through the solution for 10 min and then irradiated under an atmosphere of argon. The progress of the reactions was monitored by GC or 1 H NMR analysis of aliquot samples.

(1R,4S,5S)- and (1S,4S,5R)-4-Hydroxymethyl-3-oxabicyclo[3.2.0]hept-6-en-2-one (13 and 14). Method A. A solution of (5S)-4-hydroxymethyl-2(5H)-furanone, 4a (357 mg, 3.1 mmol), and (Z)-1,2-dichloroethylene (1.18 mL, 15.6 mmol) in acetonitrile (280 mL) was irradiated through a Quartz filter for 5 h at -20 °C. Evaporation of the solvent and column chromatography (from hexane-EtOAc 10:1 to EtOAc) afforded a diastereomeric mixture of the dichlorocyclobutane derivatives. The resulting crude was dissolved in (80%) aqueous EtOH (4 mL), and activated Zn dust (4.0 g, 61.2 mmol) was added. The mixture was irradiated under pressure in a focused microwave reactor at 100 °C for 15 min. After cooling, the reaction mixture was filtered through Celite. The solid was washed several times with EtOH and EtOAc. Evaporation of the solvent gave a residue, which was subjected to column chromatography (hexane-EtOAc 1:2) to afford a mixture of the two diastereomers 13 and 14 (328 mg, 2.34 mmol, 75% yield) in a ratio (90:10). A further column chromatography (from hexane-EtOAc 2:1 to hexane-EtOAc 1:1) allowed us to isolate pure 13 (262 mg, 1.87 mmol, 65% yield) as a colorless oil.

Method B. A solution of 2(5H)-furanone 4a (112 mg, 0.98 mmol) in freshly distilled acetone (65 mL) saturated with acetylene was irradiated through a Pyrex filter for 5.5 h at -20 °C. Evaporation of the solvent and column chromatography (hexane—EtAcO 1:1) afforded a 65:35 mixture of the cycloadducts 13 and 14 (60 mg, 0.43 mmol, 44% yield) and some unreacted 4a (19 mg, 0.17 mmol, 17%). Repeated column chromatography (from hexane—EtAcO 3:1 to hexane—EtAcO 1:1) allowed us to isolate 13 and an enriched fraction of 14.

13: IR (ATR) 3422, 3056, 2939, 2874, 1737, 1171 cm $^{-1}$; 1 H NMR (360 MHz, CDCl₃) δ 6.34 (d, $J_{6,7}$ = 2.6 Hz, 1H, H-6), 6.26 (d, $J_{7,6}$ = 2.6 Hz, 1H, H-7), 4.45 (ddd, $J_{4,8}$ = 3.6 Hz, $J_{4,8}$ = 3.6 Hz, $J_{4,5}$ = 1.7 Hz, 1H, H-4), 3.86 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 3.6 Hz, 1H, H-8), 3.67 (d, $J_{1,5}$ = 3.5 Hz, 1H, H-1), 3.62 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 3.6 Hz, 1H, H-8), 3.48 (dd, $J_{5,1}$ = 3.5 Hz, $J_{5,4}$ = 1.7 Hz, 1H, H-5), 2.99 (br s, 1H, OH); 13 C NMR (100 MHz, CDCl₃) δ 176.1 (C=O, C-2), 141.2 (CH, C-6), 138.6 (CH, C-7), 79.5 (CH, C-4), 64.4 (CH₂, C-8), 47.9 (CH, C-1), 44.1 (CH, C-5). HRMS (ESI+) calcd for $[C_7H_8O_3+H]^+$ 141.0552, found 141.0554.

14: 1 H NMR (250 MHz, CDCl₃) δ 6.44 (t, J = 2.6 Hz, 1H, H-6), 6.39 (t, J = 2.2 Hz, 1H, H-7), 4.54 (ddd, $J_{4,8}$ = 4.3 Hz, $J_{4,8}$ = 4.3 Hz, $J_{4,5}$ = 4.3 Hz, 1H, H-4), 3.95 – 3.45 (m, 4H, H-1, H-5, H-8), 1.94 (br s, 1H, OH); 13 C NMR (62.5 MHz, CDCl₃) δ 176.1 (C=O, C-2), 139.3 (CH, C-6), 138.7 (CH, C-7), 78.6 (CH, C-4), 62.6 (CH₂, C-8), 43.5 (CH, C-1), 40.1 (CH, C-5).

(15,45,5R)- and (1R,45,5S)-1-Fluoro-4-hydroxymethyl-3-oxabicyclo-[3.2.0]heptan-2-one (15 and 16). A solution of the 3-fluoro-2(5H)-furanone 4b (135 mg, 1.02 mmol) in freshly distilled acetone (65 mL) saturated with ethylene was irradiated through a Pyrex filter for 3.5 h at −20 °C. Evaporation of the solvent and column chromatography (hexane−EtOAc 3:1) afforded a 81:19 mixture of the cycloadducts 15 and 16 (131 mg, 0.81 mmol, 80% yield). Repeated column chromatography (from hexane−EtOAc 5:1 to hexane−EtOAc 3:1) allowed us to isolate pure 15 (101 mg, 0.63 mmol, 62% yield) as a white solid and an enriched fraction of 16.

15: Mp 99–102 °C (from EtOAc); $[\alpha]_D$ –36.0 (c 1.3, CHCl₃); IR (ATR) 3331, 2959, 1780, 1766, 1037 cm⁻¹; 1 H NMR (360 MHz, CDCl₃) δ 4.45 (ddd, $J_{4,8}$ = 4.4 Hz, $J_{4,F}$ = 3.9 Hz, $J_{4,8}$ = 3.4 Hz, 1H, H-4), 3.83 (dd, J_{gem} = 12.3 Hz, $J_{8,4}$ = 3.4 Hz, 1H, H-8), 3.68 (dd, J_{gem} = 12.3 Hz, $J_{8,4}$ = 3.4 Hz, 1H, H-8), 3.68 (dd, J_{gem} = 12.3 Hz, $J_{8,4}$ = 4.4 Hz, 1H, H-8), 3.20 (ddd, J = 17.2 Hz, J = 10.6 Hz, J = 6.9 Hz, 1H, H-5), 2.61 (m, 1H, H-7exo), 2.50 (m, 1H, H-7endo), 2.36 (m, 1H, H-6exo), 2.22 (br s, 1H, OH), 1.54 (m, 1H, H-6); 13 C NMR (90 MHz, CDCl₃) δ 173.9 (d, $^2J_{C-F}$ = 25.9 Hz, C=O), 90.1 (d, $^1J_{C-F}$ = 243.4 Hz, C, C-1), 84.1 (s, CH, C-4), 63.6 (s, CH₂, C-8), 41.9 (d, $^2J_{C-F}$ = 17.7 Hz, CH, C-5), 28.8 (d, $^2J_{C-F}$ = 23.8 Hz, CH, C-7), 15.4 (d, $^3J_{C-F}$ = 16.1 Hz, CH, C-6); 19 F NMR (235 MHz, CDCl₃) δ –150.6 to –150.8 (m). Anal. Calcd for (C_7 H₉FO₃): C, 52.50; H, 5.66. Found: C, 52.62; H, 5.72.

16: 1 H NMR (250 MHz, CDCl₃) δ 4.37 (ddd, $J_{4,8}$ = 7.3 Hz, $J_{4,5}$ = 4.9 Hz, $J_{4,8}$ = 4.6 Hz, 1H, H-4), 3.91 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 7.3 Hz, 1H, H-8), 3.75 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 4.6 Hz, 1H, H-8), 3.35 (m, 1H, H-5), 2.57 (m, 1H, H-7), 2.51 (m, 1H, H-7), 2.10 (m, 1H, H-6), 1.75 (br s, 1H, OH), 1.65 (m, 1H, H-6); 13 C NMR (62.5 MHz, CDCl₃) δ 173.0 (d, $^{2}J_{C-F}$ = 26.1 Hz, C=O), 90.5 (d, $^{1}J_{C-F}$ = 243.8 Hz, C, C-1), 80.7 (s, CH, C-4), 61.6 (s, CH₂, C-8), 42.6 (d, $^{2}J_{C-F}$ = 18.8 Hz, CH, C-5), 28.3 (d, $^{2}J_{C-F}$ = 21.8 Hz, CH, C-7), 9,8 (d, $^{3}J_{C-F}$ = 16.3 Hz, CH, C-6); 19 F NMR (235 MHz, CDCl₃) δ -150.8 to -151.0 (m).

(15,45,5R)-1-Chloro-4-hydroxymethyl-3-oxabicyclo[3.2.0]heptan-2-one (17) and (6R,7R)- and (6R,7S)-7-Chloro-6-hydroxymethyl-5-oxaspiro[2.4]heptan-4-one (23 and 24). A solution of the 3-chloro-2(5H)-furanone 4c (148 mg, 1.00 mmol) in freshly distilled acetone (65 mL) saturated with ethylene was irradiated through a Pyrex filter for 4.5 h at -20 °C. Evaporation of the solvent and column chromatography (hexane—EtAcO 3:1) delivered a 72:11:17 mixture of 17, 23, and 24 (148 mg, 0.83 mmol, 85% yield). Repeated column chromatography (from hexane—EtAcO 5:1 to hexane—EtAcO 3:1) allowed us to isolate 17 (102 mg, 0.57 mmol, 58% yield), 23 (14 mg, 0.08 mmol, 85% yield), and 24 (22 mg, 0.12 mmol, 13% yield) as colorless oils.

17: $[\alpha]_D$ +12.7 (c 1.1, CHCl₃); IR (ATR) 3400, 2950, 2872, 1766, 1223 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 4.46 (dd, $J_{4,8}$ = 4.9 Hz, $J_{4,8}$ = 4.7 Hz, 1H, H-4), 3.76 (m, 2H, H-8), 3.22 (ddd, $J_{5,6}$ = 7.0 Hz, $J_{5,6}$ = 1.7 Hz, $J_{5,4}$ = 0.5 Hz, 1H, H-5), 2.67 (m, 1H, H-7), 2.60 (m, 1H, H-7), 2.51 (m, 1H, H-6exo), 2.49 (br s, 1H, OH), 1.88 (m, 1H, H-6endo); ¹³C NMR (62.5 MHz, CDCl₃) δ 175.5 (C=O, C-2), 84.4 (CH, C-4), 63.1 (CH₂, C-8), 59.5 (C, C-1), 45.7 (CH, C-5), 33.4 (CH₂, C-7), 20.2 (CH₂, C-6). HRMS (ESI+) calcd for $[C_7H_9ClO_3 + Na]^+$ 199.0132, found 199.0129.

23: Mp 39–41 °C (from EtOAc); $[\alpha]_{\rm D}$ +172.2 (c 1.8, CHCl₃); IR (ATR) 3432, 2970, 1742, 1149, 1044 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 4.82 (ddd, $J_{6,8}$ = 6,7 Hz, $J_{6,8}$ = 5.0 Hz, $J_{6,7}$ = 5.0 Hz, 1H, H-6), 4.50 (d, $J_{7,6}$ = 5.0 Hz, 1H, H-7), 4.14 (dd, $J_{\rm gem}$ = 12.3 Hz, $J_{8,6}$ = 6.7 Hz, 1H, H-8), 4.00 (dd, $J_{\rm gem}$ = 12.3 Hz, $J_{8,6}$ = 5.0 Hz, 1H, H-8), 3.18 (br s, 1H, OH), 1.59–1.30 (m, 4H, H-1, H-2); ¹³C NMR (90 MHz, CDCl₃) δ 176.2 (C=O, C-4), 80.0 (CH, C-6), 62.8 (CH, C-7), 62.2 (CH₂, C-8),

28.6 (C, C-3), 19.0/13.5 (2CH₂, C-1, C-2). HRMS (ESI+) calcd for $[C_7H_9ClO_3 + Na]^+$ 199.0132, found 199.0132.

24: $[\alpha]_D$ –69.7 (*c* 2.54, CHCl₃); IR (ATR) 3423, 2934, 1755, 1112, 1055 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 4.73 (ddd, $J_{6,8}$ = 3.5 Hz, $J_{6,8}$ = 3.3 Hz, $J_{6,7}$ = 3.3 Hz, 1H, H-6), 4.49 (d, $J_{7,6}$ = 3.3 Hz, 1H, H-7), 3.95 (dd, J_{gem} = 12.7 Hz, $J_{8,6}$ = 3.3 Hz, 1H, H-8), 3.79 (dd, J_{gem} = 12.7 Hz, $J_{8,6}$ = 3.5 Hz, 1H, H-8), 3.32 (br s, 1H, OH), 1.48–1.27 (m, 4H, H-1, H-2); ¹³C NMR (90 MHz, CDCl₃) δ 177.6 (C=O, C-4), 86.4 (CH, C-6), 61.6 (CH₂, C-8), 59.8 (CH, C-7), 28.1 (C, C-3), 19.2/15.6 (2CH₂, C-1, C-2). HRMS (ESI+) calcd for $[C_7H_9ClO_3 + Na]^+$ 199.0132, found 199.0134.

(15,45,5R)- and (1R,45,5S)-1-Fluoro-4-hydroxymethyl-3-oxabicyclo-[3.2.0]hept-6-en-2-one (**19** and **20**) and (1R,25,5R,65,9S)-6-Fluoro-9-hydroxymethyl-8-oxatricyclo[4.3.0.0^{2,5}]non-3-en-7-one (**25**). A solution of lactone **4b** (130 mg, 0.98 mmol) in freshly distilled acetone (65 mL) saturated with acetylene was irradiated through a Pyrex filter for 5.5 h at -20 °C. Evaporation of the solvent and column chromatography (from hexane—EtOAc 6:1 to hexane—EtOAc 4:1) afforded the following fractions: (i) a 95:5 mixture of the cyclobutene derivatives **19** and **20** (44 mg, 0.27 mmol, 28% yield) as a colorless oil; (ii) the tricyclic derivative **25** (27 mg, 0.15 mmol, 15%) as a colorless oil; and (iii) some unreacted **4b** (45 mg, 0.34 mmol, 35%).

19: IR (ATR) 3404, 2937, 2878, 1761, 1075 cm⁻¹; 1 H NMR (360 MHz, CDCl₃) δ 6.57 (ddd, $J_{6,F}$ = 11.4 Hz, $J_{6,7}$ = 2.8 Hz, $J_{6,5}$ = 0.6 Hz, 1H, H-6), 6.43 (ddd, $J_{7,6}$ = 2.7 Hz, $J_{7,F}$ = 1.3 Hz, $J_{7,5}$ = 0.6 Hz, 1H, H-7), 4.38 (dddd, $J_{4,F}$ = 4.5 Hz, $J_{4,8}$ = 3.2 Hz, $J_{4,8}$ = 2.9 Hz, $J_{4,5}$ = 1.5 Hz, 1H, H-4), 3.94 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 2.9 Hz, 1H, H-8), 3.71 (dd, J_{gem} = 12.2 Hz, $J_{8,4}$ = 3.2 Hz, 1H, H-8), 3.59 (dddd, $J_{5,F}$ = 2.6 Hz, $J_{5,4}$ = 1.5 Hz, $J_{5,6}$ = 0.6 Hz, $J_{5,7}$ = 0.6 Hz, 1H, H-5); 13 C NMR (90 MHz, CDCl₃) δ 171.0 (d, $^{2}J_{C-F}$ = 28.9 Hz, C=O, C-2), 141.3 (d, $^{3}J_{C-F}$ = 15.6 Hz, CH, C-6), 138.6 (d, $^{2}J_{C-F}$ = 26.2 Hz, CH, C-7), 91.6 (d, $^{1}J_{C-F}$ = 248.2 Hz, C, C-1), 78.3 (s, CH, C-4), 63.5 (s, CH₂, C-8), 49.6 (d, $^{2}J_{C-F}$ = 16.4 Hz, CH, C-5); 19 F NMR (235 MHz, CDCl₃) δ -165.6 to -165.9 (m). HRMS (ESI+) calcd for $[C_{7}H_{7}FO_{3}$ + Na]⁺ 181.0199, found 181.0193 (mixture 19 and 20).

20: ¹H NMR (250 MHz, CDCl₃) δ 6.71 (ddd, $J_{6,F}$ = 7.9 Hz, $J_{6,7}$ = 2.9 Hz, $J_{6,5}$ = 0.7 Hz, 1H, H-6), 6.23 (br d, $J_{7,6}$ = 2.9 Hz, 1H, H-7), 4.95 (m, 1H, H-4), 3.98 (dd, J_{gem} = 12.3 Hz, $J_{8,4}$ = 2.8 Hz, 1H, H-8), 3.79 (dd, J_{gem} = 12.3 Hz, $J_{8,4}$ = 4.9 Hz, 1H, H-8), 3.60 (m, 1H, H-5).

25: $[\alpha]_D + 28.9$ (c 0.44, CHCl₃); IR (ATR) 3399, 2940, 1762, 1225, 1063 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 6.45 (m, 1H, H-3), 6.19 (ddd, J = 2.6 Hz, J_{4,2} = 2.1 Hz, J = 0.7 Hz, 1H, H-4), 4.34 (dddd, J_{9,F} = 6.4 Hz, J_{9,10} = 4.3 Hz, J_{9,10} = 3.2 Hz, J_{9,11} = 1.9 Hz, 1H, H-9), 3.84 (dd, J_{gem} = 12.3 Hz, J_{10,9} = 3.2 Hz, 1H, H-10), 3.78 (m, 1H, H-5), 3.68 (dd, J_{gem} = 12.3 Hz, J_{10,9} = 4.3 Hz, 1H, H-10), 3.56 (dddd, J_{2,1} = 8.0 Hz, J = 5.9 Hz, J = 3.4 Hz, J_{2,4} = 2.1 Hz, 1H, H-2), 3.09 (dddd, J_{1,F} = 16.9 Hz, J_{1,2} = 8.0 Hz, J_{1,9} = 1.9 Hz, J_{1,3} = 1.0 Hz, 1H, H-5), 2.19 (br s, 1H, OH); ¹³C NMR (90 MHz, CDCl₃) δ 171.8 (d, 2J _{C-F} = 25.2 Hz, C=O, C-7), 140.7 (d, 4J _{C-F} = 3.3 Hz, CH, C-3), 137.8 (d, 3J _{C-F} = 4.2 Hz, CH, C-4), 95.9 (d, 1J _{C-F} = 238.9 Hz, C, C-6), 82.3 (d, 3J _{C-F} = 3.9 Hz, CH, C-9), 64.0 (s, CH₂, C-10), 49.9 (d, 2J _{C-F} = 49.9 Hz, CH, C-5), 38.4 (d, 3J _{C-F} = 4.6 Hz, CH, C-2), 37.9 (d, 2J _{C-F} = 16.9 Hz, CH₂, C-1); ¹⁹F NMR (235 MHz, CDCl₃) δ – 168.0 to –168.2 (m). HRMS (ESI+) calcd for [C₉H₉FO₃ + Na]⁺ 207.0428, found 207.0430.

(15,45,5R)-1-Chloro-4-hydroxymethyl-3-oxabicyclo[3.2.0]hept-6-en-2-one (**21**) and (1R,25,5R,65,9S)-6-Chloro-9-hydroxymethyl-8-oxatricyclo[4.3.0.0^{2,5}]non-3-en-7-one (**26**). A solution of lactone **4c** (149 mg, 1.00 mmol) in freshly distilled acetone (65 mL) saturated with acetylene was irradiated through a Pyrex filter for 4.5 h at -20 °C. Evaporation of the solvent and column chromatography (hexane—EtAcO 3:1) afforded the cycloadduct **21** (29 mg, 0.17 mmol, 17% yield) as a colorless oil, the tricyclic derivative **26** (12 mg, 0.06 mmol, 6% yield) as a colorless oil, and some unreacted **4c** (36 mg, 0.24 mmol, 24%).

21: $[\alpha]_D$ -71.1 (*c* 2.54, CHCl₃); IR (ATR) 3405, 2933, 1755, 957 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 6.50 (dd, $J_{6,7}$ = 2.6 Hz,

 $J_{6,5} = 0.6$ Hz, 1H, H-6), 6.37 (d, $J_{7,6} = 2.6$ Hz, 1H, H-7), 4.44 (ddd, $J_{4,8} = 3.5$ Hz, $J_{4,8} = 3.0$ Hz, $J_{4,5} = 1.5$ Hz, 1H, H-4), 3.93 (dd, $J_{\rm gem} = 12.0$ Hz, $J_{8,4} = 3.0$ Hz, 1H, H-8), 3.72 (dd, $J_{\rm gem} = 12.0$ Hz, $J_{8,4} = 3.5$ Hz, 1H, H-8), 3.60 (dd, $J_{5,4} = 1.0$ Hz, $J_{5,6} = 0.6$ Hz, 1H, H-5), 2.08 (br s, 1H, OH); 13 C NMR (90 MHz, CDCl₃) δ 171.5 (C=O, C-2), 140.5 (CH, C-7), 139.7 (CH, C-6), 78.1 (CH, C-4), 64.0 (C, C-1), 63.7 (CH₂, C-8), 53.5 (CH, C-5). HRMS (ESI+) calcd for $[C_7H_7ClO_3+Na]^+$ 196.9976, found 196.9972.

HRMS (ESI+) calcd for $[C_7H_7C_1O_3+Na]^T$ 196.99%, found 196.99%. **26**: IR (ATR) 3395, 2942, 2860, 1765, 1080 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 6.49 (dd, $J_{3,5}$ = 2.6 Hz, $J_{3,4}$ = 2.4 Hz, 1H, H-3), 6.35 (dd, $J_{4,3}$ = 2.4 Hz, $J_{4,2}$ = 2.1 Hz, 1H, H-4), 4.39 (ddd, $J_{9,10}$ = 4.6 Hz, $J_{9,10}$ = 3.4 Hz, 1H, H-10), 3.75 (dd, $J_{5,2}$ = 2.8 Hz, $J_{5,3}$ = 2.6 Hz, 1H, H-5), 3.68 (dd, J_{gem} = 12.3 Hz, $J_{10,9}$ = 3.4 Hz, 1H, H-10), 3.65 (ddd, $J_{2,1}$ = 7.9 Hz, $J_{2,5}$ = 2.7 Hz, $J_{2,4}$ = 2.1 Hz, 1H, H-2), 3.15 (dd, $J_{1,2}$ = 7.9 Hz, $J_{1,9}$ = 1.8 Hz, 1H, H-5), 1.90 (br s, 1H, OH); ¹³C NMR (125 MHz, CDCl₃) δ 173.3 (C=O, C-7), 140.2/139.7 (2CH, C-3, C-4), 77.2 (CH, C-9), 64.0 (CH₂, C-10), 63.1 (C, C-6), 53.4 (CH, C-1), 43.6 (CH, C-5), 40.1 (CH, C-2). HRMS (ESI+) calcd for $[C_9H_9ClO_3 + Na]^+$ 223.0132, found 223.0135.

(1R,4S,5S)-4-tert-Butyldimethylsilyloximethyl-3-oxabicyclo[3.2.0] hept-6-en-2-one (13a). To an ice-cooled solution of 13 (417 mg, 2.98 mmol) in CH₂Cl₂ (20 mL) were added imidazole (283 mg, 4.16 mmol) and tert-butyl dimethylsilyl chloride (584 mg, 3.87 mmol). The mixture was allowed to stir 3 h at room temperature and then was diluted with CH₂Cl₂ (10 mL) and washed with water (10 mL). The organic layer was dried with anhydrous Na2SO4, filtered, and concentrated to dryness. The reaction crude was purified by column chromatography (hexane— EtOAc, 10:1) to give 13a (585 mg, 2.30 mmol, 77% yield) as a white solid: mp 49–50 °C (from EtOAc–pentane); [α]_D –221.9 (\emph{c} 0.55, CHCl₃); MS (ESI+) 255 ($[M + H]^+$, 28), 277 ($[M + Na]^+$, 50); IR (ATR) 2957, 2926, 2856, 1750, 1250, 1171, 1111 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 6.32 (ddd, $J_{6,7}$ = 2.7 Hz, $J_{6,5}$ = 0.7 Hz, $J_{6,1}$ = 0.7 Hz, 1H, H-6), 6.26 (dd, $J_{7,6}$ = 2.7 Hz, $J_{7,1}$ = 0.8 Hz, 1H, H-7), 4.39 (dddd, $J_{4,8}$ = 2.9 Hz, $J_{4,8} = 2.2$ Hz, $J_{4,5} = 1.3$ Hz, $J_{4,1} = 0.6$ Hz, 1H, H-4), 3.83 (dd, $J_{gem} =$ 10.9 Hz, $J_{4,8} = 2.9 \text{ Hz}$, 1H, 1H-8), 3.63 (dd, $J_{\text{gem}} = 10.9 \text{ Hz}$, $J_{4,8} = 2.2 \text{ Hz}$, 1H, H-8), 3.59 (dddd, $J_{1,5} = 3.5$ Hz, $J_{1,6} = 0.7$ Hz, $J_{1,4} = 0.7$ Hz, $J_{1,7} = 0.7$ Hz, 1H, H-1), 3.48 (ddd, $J_{5,1}$ = 3.5 Hz, $J_{5,4}$ = 1.3 Hz, $J_{5,6}$ = 0.6 Hz, 1H, H-5), 0.86 (s, 9H, $C(CH_3)_3$), 0.05 (s, 3H, CH_3Si), 0.04 (s, 3H, CH_3Si); ¹³CNMR (62.5 MHz, CDCl₃) δ 175.5 (C=O), 140.9 (CH, C-6), 138.9 (CH, C-7), 78.6 (CH, C-4), 65.0 (CH₂, C-8), 48.2 (CH, C-1), 44.6 (CH, C-5), 25.7 (CH₃, C(CH₃)₃), 18.0 (C, C(CH₃)₃), -5.7 (CH₃Si), -5.7 (CH₃Si). Anal. Calcd for (C₁₃H₂₂O₃Si): C, 61.38; H, 8.72. Found: C, 61.47: H. 8.74.

(1R,2RS,4S,5S)-2-Acetyloxy-4-tert-butyldimethylsilyloxymethyl-3oxabicyclo[3.2.0]hept-6-ene (27). To a solution of 13a (398 mg, 1.56 mmol) in dry toluene (45 mL) at -78 °C was added dropwise a 1.0 M solution of DIBAL-H in toluene (7.8 mL, 7.8 mmol) over a period of 30 min under nitrogen atmosphere. After 2 h of stirring at — 78 °C, the reaction mixture was quenched by the slow addition of methanol (2 mL) and allowed to warm to room temperature. EtOAc (4 mL) and a saturated NaHCO₃ solution (1 mL) were added. After 20 min of vigorous stirring, anhydrous Na₂SO₄ (6.6 g) was added. The suspension was stirred vigorously overnight, filtered through Celite, and concentrated to dryness, affording a crude reaction which was used in the next reaction without further purification. To an icecooled solution of this crude in CH₂Cl₂ (15 mL), pyridine (2.5 mL) and acetic anhydride (2.2 mL) were successively added dropwise. The reaction mixture was stirred at room temperature overnight. Then, the reaction was washed with HCl 5% (2 × 10 mL), saturated NaHCO₃ solution (3 × 10 mL), and brine (10 mL). The organic layer was dried with anhydrous Na2SO4, filtered, and evaporated to dryness. The crude residue was purified by column chromatography (hexane-EtOAc, 10:1) to give 27 (385 mg, 1.29 mmol, 83% yield for the 2 steps) as a colorless oil: [α]_D -94.2 (c 0.55, CHCl₃); MS (ESI+) 321 $([M + Na]^+, 100)$; IR (ATR) 2958, 2930, 2856, 1740,

1226 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 6.13 (m, 3H, H-6, H-7, H-2), 4.06 (dddd, $J_{4,8}$ = 9.0 Hz, $J_{4,8}$ = 5.5 Hz, $J_{4,5}$ = 0.9 Hz, $J_{4,1}$ = 0.5 Hz, 1H, H-4), 3.64 (dd, $J_{\rm gem}$ = 9.9 Hz, $J_{8,4}$ = 5.5 Hz, 1H, H-8), 3.49 (dd, $J_{\rm gem}$ = 9.9 Hz, $J_{8,4}$ = 9.0 Hz, 1H, H-8), 3.48 (m, 1H, H-1), 3.41 (m, 1H, H-5), 2.02 (s, 3H, CH₃CO), 0.90 (s, 9H, C(CH₃)₃), 0.07 (s, 6H, 2 × CH₃Si); ¹³C NMR (62.5 MHz, CDCl₃) δ 170.0 (C=O), 140.5 (CH, C-6), 136.9 (CH, C-7), 97.7 (CH, C-2), 80.8 (CH, C-4), 64.6 (CH₂, C-8), 53.4 (CH, C-1), 48.6 (CH, C-5), 25.8 (CH₃, C(CH₃)₃), 21.5 (CH₃, CH₃CO), 18.3 (C, C(CH₃)₃), -5.3 (CH₃, CH₃Si), -5.4 (CH₃, CH₃Si). Anal. Calcd for (C₁₅H₂₆O₄Si): C, 60.37; H, 8.78. Found: C, 60.25; H, 8.53.

(1'R,2'S,4'S,5'S)- and (1'R,2'R,4'S,5'S)-6-Chloro-9-(4-tert-butyldimethylsilyloxymethyl-3-oxabicyclo[3.2.0]hept-6-en-2-yl)-9H-purine (28 and 29). BSA (1.06 mL, 4.13 mmol) was added to a suspension of 6-chloropurine (319 mg, 2.06 mmol) in dry acetonitrile (15 mL) under an argon atmosphere. The reaction was allowed to stir for 20 min and cooled to 0 °C. Then, a solution of 27 (411 mg, 1.38 mmol) in dry acetonitrile (4 mL) and TMSOTf (346 µL, 1.79 mmol) was successively added, and the reaction mixture was stirred at room temperature for 2 h. CH₂Cl₂ (25 mL) was added, and the reaction was quenched with an aqueous saturated solution of NaHCO3 (10 mL). The two layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 15 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and evaporated to dryness. The crude residue was purified by column chromatography (hexane-EtOAc, 6:1) to give the α-anomer 28 (220 mg, 0.56 mmol, 41% yield) as a white solid and the β -anomer 29 (243 mg, 0.62 mmol, 45% yield) as a white solid.

28: Mp 125–126 °C (from EtOAc–pentane); $[\alpha]_D$ –95.0 (c 0.80, CHCl₃); IR (ATR) 3107, 3049, 2952, 2930, 2852, 1591, 1559, 1335, 1214 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.70 (s, 1H, H-2), 8.42 (s, 1H, H-8), 6.49 (d, $J_{2',1'}$ = 5.4 Hz, 1H, H-2'), 6.37 (d, $J_{6',7'}$ = 2.8 Hz, 1H, H-6'), 5.64 (d, $J_{7',6'}$ = 2.8 Hz, 1H, H-7'), 4.17 (m, 1H, H-4'), 3.90 (m, 1H, H-1'), 3.84 (dd, J_{gem} = 10.8 Hz, $J_{8',4'}$ = 3.5 Hz, 1H, H-8'), 3.67 (dd, J_{gem} = 10.8 Hz, $J_{8',4'}$ = 3.0 Hz, 1H, H-8'), 3.58 (m, J = 3.5 Hz, 1H, H-5'), 0.94 (s, 9H, C(CH₃)₃), 0.10 (s, 3H, CH₃Si), 0.09 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 151.9 (CH, C-2), 150.9/150.6 (C, C-4/C-6), 143.6 (CH, C-8), 142.1 (CH, C-6'), 134.6 (CH, C-7'), 131.6 (C, C-5), 83.9 (CH, C-2'), 77.4 (CH, C-4'), 66.2 (CH₂, C-8'), 51.3 (CH, C-1'), 49.3 (CH, C-5'), 25.8 (CH₃, C(CH₃)₃), 18.1 (C, C(CH₃)₃), -5.5 (CH₃, CH₃Si), -5.6 (CH₃, CH₃Si). Anal. Calcd for (C₁₈H₂₅ClN₄O₂Si): C, 55.02; H, 6.41; N, 14.26. Found: C, 55.25; H, 6.47; N, 14.07.

29: Mp 79–80 °C (from EtOAc–pentane); $[\alpha]_D$ –18.7 (c 0.75, CHCl₃); IR (ATR) 3046, 2854, 2926, 2853, 1590, 1552, 1250. 1073 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.74 (s, 1H, H-2), 8.48 (s, 1H, H-8), 6.37 (dt, $J_{7',6'}$ = 2.8 Hz, $J_{7',1'}$ = 0.7 Hz, $J_{7',5'}$ = 0.7 Hz, 1H, H-7'), 6.32 (m, $J_{6',7'}$ = 2.8 Hz, 1H, H-6'), 6.19 (s, 1H, H-2'), 4.44 (m, $J_{1',5'}$ = 3.6 Hz, 1H, H-1'), 4.32 (m, 1H, H-4'), 3.61 (m, 1H, H-5'), 3.52 (dd, J_{gem} = 11.0 Hz, $J_{8',4'}$ = 5.4 Hz, 1H, H-8'), 3.46 (dd, J_{gem} = 11.0 Hz, $J_{8',4'}$ = 6.2 Hz, 1H, H-8'), 0.81 (s, 9H, C(CH₃)₃), –0.02 (s, 3H, CH₃Si), –0.05 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 151.6 (CH, C-2), 151.1/150.9 (C, C-4/C-6), 144.3 (CH, C-8), 141.1 (CH, C-6'), 137.6 (CH, C-7'), 132.5 (C, C-5), 86.6 (CH, C-2'), 82.7 (CH, C-4'), 64.7 (CH₂, C-8'), 53.5 (CH, C-1'), 49.6 (CH, C-5'), 25.8 (CH₃, C(CH₃)₃), 18.2 (C, C(CH₃)₃), –5.5 (CH₃, 2 × CH₃Si). Anal. Calcd for (C₁₈H₂₅ClN₄O₂Si): C, 55.02; H, 6.41; N, 14.26. Found: C, 55.07; H, 6.46; N, 14.21.

(1'R,2'R,4'S,5'S)-6-Chloro-9-(4-hydroxymethyl-3-oxabicyclo-[3.2.0]hept-6-en-2-yl)-9H-purine (**30**). To a solution of **28** (100 mg, 0.25 mmol) in dry THF (3 mL) was added a 1.0 M solution of TBAF in THF (0.51 mL, 0.51 mmol). The reaction was allowed to stir for 2 h, and the solvent was removed. The residue was purified by column chromatography (EtOAc—hexane, 4:1) to give **30** (53 mg, 0.21 mmol, 75% yield) as a white solid: mp 143—145 °C (from EtOAc—pentane); $[\alpha]_D$ —127.1 (c 1.40, MeOH); IR (ATR) 3345, 3117, 2954, 1592, 1562, 1397,

1203 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.66 (s, 1H, H-2), 8.43 (s, 1H, H-8), 6.38 (d, $J_{6',7'}$ = 2.9 Hz, 1H, H-6'), 6.35 (d, $J_{2',1'}$ = 5.4 Hz, 1H, H-2'), 5.66 (d, $J_{7',6'}$ = 2.9 Hz, 1H, H-7'), 4.26 (ddd, $J_{4',8'}$ = 5.3 Hz, $J_{4',8'}$ = 4.4 Hz, $J_{4',5'}$ = 0.7 Hz, 1H, H-4'), 3.91 (m, 1H, H-1'), 3.74 (m, $J_{8',4'}$ = 5.8 Hz, $J_{8',4'}$ = 4.3 Hz, 2H, H-8'), 3.50 (m, $J_{5',1'}$ = 3.6 Hz, 1H, H-5'), 3.07 (br s, 1H, OH); ¹³C NMR (90 MHz, CDCl₃) δ 151.8 (CH, C-2), 150.8/150.7 (C, C-4/C-6), 143.7 (CH, C-8), 142.0 (CH, C-6'), 134.5 (CH, C-7'), 131.4 (C, C-5), 82.9 (CH, C-2'), 77.7 (CH, C-4'), 64.0 (CH₂, C-8'), 50.7 (CH, C-1'), 48.7 (CH, C-5').

(1'R,2'R,4'S,5'S)-6-Chloro-9-(4-hydroxymethyl-3-oxabicyclo[3.2.0]hept-6-en-2-yl)-9H-purine (31). To a solution of 29 (99 mg, 0.25 mmol) in THF (3 mL) was added a 1.0 M solution of TBAF in THF (0.50 mL, 0.50 mmol). The reaction was allowed to stir for 2 h, and the solvent was removed. The residue was purified by column chromatography (hexane-EtOAc, 1:4) to give 31 (59 mg, 0.21 mmol, 84% yield) as a white solid: mp 128–129 °C (from EtOAc–pentane); $[\alpha]_D$ –24.0 (c 1.00, CHCl₃); IR (ATR) 3287, 3112, 3073, 2952, 2900, 2868, 1590, 1555, 1204 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.72 (s, 1H, H-2), 8.54 (s, 1H, H-8), 6.34 (s, 2H, H-6', H-7'), 6.25 (s, 1H, H-2'), 4.39 (ddd, $J_{4',8'} = 5.8$ Hz, $J_{4',8'} = 3.6$ Hz, $J_{4',5'} = 1.9 Hz$, 1H, H-4'), 4.35 (m, 1H, H-1'), 3.73 (m, 1H, H-5'), 3.66(dd, $J_{\text{gem}} = 11.6 \text{ Hz}$, $J_{8',4'} = 3.6 \text{ Hz}$, 1H, H-8'), 3.52 (dd, $J_{\text{gem}} = 11.6 \text{ Hz}$, $J_{8',4'}$ = 5.8 Hz, 1H, H-8'), 3.32 (br s, 1H, OH); ¹³C NMR (62.5 MHz, CDCl₃) δ 151.7 (CH, C-2), 151.1/150.6 (C, C-4/C-6), 144.8 (CH, C-8), 141.2 (CH, C-6'), 137.4 (CH, C-7'), 131.7 (C, C-5), 86.0 (CH, C-2'), 82.8 (CH, C-4'), 63.9 (CH₂, C-8'), 53.5 (CH, C-1'), 49.4 (CH, C-5').

(1'R,2'S,4'S,5'S)-9-(4-Hydroxymethyl-3-oxabicyclo[3.2.0]hept-6en-2-yl)adenine (32). A solution of 30 (19 mg, 0.068 mmol) and saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 36 h. After cooling to room temperature the solvent was removed under vacuum, and the residue was purified by column chromatography (CH_2Cl_2 : MeOH, 9:1) to afford 32 (17 mg, 0.066 mmol, 97% yield) as a white solid: mp 164–166 °C (from ether); $[\alpha]_D$ –84.5 (c 1.0, DMSO); IR (ATR) 3359, 3326, 3191, 2933, 1650, 1602 cm⁻¹; ¹H NMR (360 MHz, CD₃OD) δ 8.24 (s, 1H, H-8), 8.19 (s, 1H, H-2), 6.39 (ddd, $J_{2',1'}$ = 5.4 Hz, 1H, H-2'), 5.70 (ddd, $J_{7',6'}$ = 2.9 Hz, $J_{7',5'}$ = 0.5 Hz, $J_{7',1'}$ = 0.5 Hz, 1H, H-7'), 4.19 (ddd, $J_{4',8'} = 4.9 \text{ Hz}, J_{4',8'} = 4.9 \text{ Hz}, J_{4'5'} = 1.0 \text{ Hz}, 1\text{H}, \text{H}-4'), 3.92 \text{ (dddd}, } J_{1',2'} = 5.1$ Hz, $J_{1',5'} = 3.6 Hz$, $J_{1',6'} = 0.6 Hz$, $J_{1',7'} = 0.6 Hz$, 1H, H-1'), 3.66 (d, $J_{8',4'} = 4.9$ Hz, 2H, H-8'), 3.53 (dddd, $J_{5',1'} = 3.6$ Hz, $J_{5',4'} = 1.1$ Hz, $J_{5',6'} = 0.6$ Hz, $J_{5',7'} = 1.1$ Hz, $J_{5',6'} = 0.6$ Hz, $J_{5',7'} = 0.6$ 0.6 Hz, 1H, H-5'); 13 C NMR (90 MHz, CD₃OD) δ 157.3 (C, C-6), 153.9 (C, C-2), 149.8 (C, C-4), 143.2 (CH, C-6'), 140.4 (CH, C-8), 136.0 (CH, C-7'), 120.0 (C, C-5), 84.4 (CH, C-2'), 79.2 (CH, C-4'), 64.7 (CH₂, C-8'), 52.3 (CH, C-1'), 50.5 (CH, C-5'). HRMS (ESI+) calcd for [(C₁₂H₁₃- N_5O_2) + H]⁺ 260.1142, found 260.1133.

(1'R,2'R,4'S,5'S)-9-(4-Hydroxymethyl-3-oxabicyclo[3.2.0]hept-6-en-2-yl)adenine (33). A solution of 31 (21 mg, 0.075 mmol) and saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 36 h. After cooling to room temperature the solvent was removed under vacuum, and the residue was purified by column chromatography (CH₂Cl₂: MeOH, 9:1). Recrystallization with MeOH gave 33 (14 mg, 0.054 mmol, 72% yield) as a white solid: mp 229–231 °C (from MeOH); $[\alpha]_D$ – 13.3 (c 0.9, DMSO); IR (ATR) 3230, 3117, 3144, 2931, 2820, 1687, 1606, 1569, 1296 cm $^{-1}$; 1 H NMR (360 MHz, CD $_{3}$ OD) δ 8.35 (s, 1H, H-8), 8.20 (s, 1H, H-2), 6.34 (ddd, $J_{7',6'}$ = 2.8 Hz, $J_{7',1'}$ = 0.8 Hz, $J_{7',5'}$ = 0.8 Hz, 1H, H-7'), 6.32 (m, $J_{6',7'}$ = 2.8 Hz, 1H, H-6'), 6.18 (s, 1H, H-2'), 4.36 (ddd, $J_{1',5'} = 3.6 \,\text{Hz}$, $J_{1',7'} = 0.8 \,\text{Hz}$, $J_{1',2'} = 0.8 \,\text{Hz}$, 1H, H-1'), 4.21 (m, 1H, H-4'), 3.68 (m, 1H, H-5'), 3.36 (d, $J_{8',4'}$ = 1.5 Hz, 1H, H-8'), 3.34 (d, $J_{8',4'}$ = 2.2 Hz, 1H, H-8'); ¹³C NMR (90 MHz, CD₃OD) δ 157.3 (C, C-6), 153.7 (CH, C-1), 150.2 (C, C-4), 142.2 (CH, C-6'), 141.3 (CH, C-8), 139.2 (CH, C-7'), 120.6 (C, C-5), 86.5 (CH, C-2'), 83.8 (CH, C-4'), 64.8 (CH₂, C-8'), 54.3 (CH, C-1'), 51.3 (CH, C-5'). HRMS (ESI+) calcd for $([(C_{12}H_{13}N_5O_2) + Na]^+)$ 282.0961, found 282.0960. (1S,4S,5R)-4-tert-Butyldimethylsilyloxymethyl-1-fluoro-3-oxabicyclo-

(15,45,5R)-4-tert-Butyldimethylsilyloxymethyl-1-fluoro-3-oxabicyclo-[3.2.0]heptan-2-one (**15a**). To a solution of **15** (410 mg, 2.56 mmol) in CH_2Cl_2 (18 mL) at 0 °C were successively added imidazole (360 mg,

5.26 mmol) and tert-butyldimethylsilyl chloride (536 mg, 3.57 mmol). The mixture was allowed to warm to room temperature and stirred for 16 h. The reaction mixture was successively washed with 5% aqueous citric acid solution (3 \times 10 mL), saturated aqueous NaHCO₃ solution (3 \times 10 mL), and brine (3 \times 10 mL). Then, the organic layer was dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness. Purification by column chromatography (hexane-diethyl ether 10:1) afforded 15a (618 mg, 2.25 mmol, 88% yield) as a white solid: mp 48-49 °C (from diethyl ether); $[\alpha]_D$ -53.0 (c 1.0, CHCl₃); IR (ATR) 2929, 1780, 1098, 1081, 827 cm⁻¹; 1 H NMR (250 MHz, CDCl₃) δ 4.37 (ddd, $J_{4,8}$ = 3.6 Hz, $J_{4,F}$ = 3.3 Hz, $J_{4,8}$ = 2.7 Hz, 1H, H-4), 3.81 (dd, J_{gem} = 11.2 Hz, $J_{8,4}$ = 3.6 Hz, 1H, H-8), 3.67 (dd, $J_{\text{gem}} = 11.2 \text{ Hz}$, $J_{8,4} = 2.7 \text{ Hz}$, 1H, H-8), 3.21 (dddd, J = 10.6 Hz, J = 10.4 Hz, J = 6.6 Hz, $J_{5,4} = 0.5$ Hz, 1H, H-5), 2.59 (m, 1H, H-7exo), 2.50 (m, 1H, H-7endo), 2.32 (m, 1H, H-6exo), 1.52 (m, 1H, H-6endo), 0.87 (s, 9H, $(CH_3)_3C$), 0.06 (s, 3H, CH_3Si), 0.05 (s, 3H, CH_3Si); ¹³C NMR (62.5 MHz, CDCl₃) δ 173.8 (d, ${}^{2}J_{C-F}$ = 25.4 Hz, C=O), 90.5 (d, ${}^{1}J_{C-F}$ = 243.2 Hz, C, C-1), 83.9 (d, ${}^{3}J_{C-F} = 2.3$ Hz, CH, C-4), 63.8 (s, CH₂, C-8), 42.4 (d, ${}^{2}J_{C-F} =$ 17.8 Hz, CH, C-5), 29.1 (d, ${}^{2}J_{C-F}$ =24.4 Hz, CH₂, C-7), 25.7 (s, CH₃, (CH₃)₃C), 18.2 (s, C, (CH₃)₃C), 15.6 (d, ${}^{3}J_{C-F}$ = 15.8 Hz, CH₂, C-6), -5.7 (s, CH₃, CH₃Si), -5.8 (s, CH₃, CH₃Si); ${}^{19}F$ NMR (235 MHz, CDCl₃) δ -150.8 to -151.0 (m). HRMS (ESI+) calcd for $[C_{13}H_{23}FO_3Si + H]^+$ 275.1473, found 275.1474.

(1S,2R,4S,5R)- and (1S,2S,4S,5R)-2-Acetyloxy-4-tert-butyldimethylsilyloxymethyl-1-fluoro-3-oxabicyclo[3.2.0]heptane (34 and 35). To a solution of **15a** (1.79 g, 6.52 mmol) in dry CH₂Cl₂ (30 mL) at −78 °C was added dropwise a 1.0 M solution of DIBAL-H in CH₂Cl₂ (9.3 mL₂ 9.3 mmol). After 2.5 h of stirring at -78 °C, the reaction mixture was quenched by the slow addition of an aqueous HNO₃ 3% solution (25 mL) and allowed to warm to room temperature. The two layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness affording a reaction crude which was used in the next reaction without further purification. To an ice-cooled solution of this crude and pyridine (7.4 mL, 92.7 mmol) in CH₂Cl₂ (50 mL) was added dropwise acetic anhydride (5.9 mL, 61.8 mmol). The reaction mixture was stirred at room temperature overnight. Then, the reaction was washed with HCl 3% $(3 \times 20 \text{ mL})$, saturated NaHCO₃ solution $(3 \times 20 \text{ mL})$, and brine $(3 \times$ 20 mL). The organic layer was dried with anhydrous Na₂SO₄, filtered, and evaporated to dryness. The resulting crude residue was purified by column chromatography (hexane-diethyl ether 10:1) to afford a 84:16 mixture of acetates 34 and 35 (1.84 g, 5.78 mmol, 89% yield for the 2 steps). A further column chromatography (from hexane-diethyl ether 20:1 to hexanediethyl ether 5:1) allowed us to isolate pure α -acetate 34 and β -acetate 35 both as colorless oils.

34: $[\alpha]_D$ +22.4 (c 1.25, CHCl₃); IR (ATR) 2929, 1752, 1222, 1105 cm⁻¹; 1 H NMR (360 MHz, CDCl₃) δ 6.31 (d, $J_{2,F}$ = 14.5 Hz, 1H, H-2), 4.14 (dddd, $J_{4,8}$ = 5.6 Hz, $J_{4,8}$ = 4.2 Hz, $J_{4,5}$ = 2.1 Hz, $J_{4,F}$ = 2.1 Hz, 1H, H-4), 3.67 (dd, J_{gem} = 10.7 Hz, $J_{8,4}$ = 4.2 Hz, 1H, H-8), 3.61 (dd, J_{gem} = 10.7 Hz, $J_{8,4}$ = 5.6 Hz, 1H, H-8), 3.01 (m, 1H, H-5), 2.40 (m, 1H, H-7endo), 2.30 (m, 1H, H-7exo), 2.15 (m, 4H, H-6exo, CH₃CO), 1.50 (m, 1H, H-6endo), 0.89 (s, 9H, (CH₃)₃C), 0.06 (s, 3H, CH₃Si), 0.05 (s, 3H, CH₃Si); 13 C NMR (90 MHz, CDCl₃) δ 169.5 (s, C=O), 102.8 (d, $^{1}J_{C-F}$ = 234.2 Hz, C, C-1), 101.0 (d, $^{2}J_{C-F}$ = 42.0 Hz, CH, C-2), 87.1 (s, CH, C-4), 64.6 (s, CH₂, C-8), 45.8 (d, $^{2}J_{C-F}$ = 18.9 Hz, CH, C-5), 25.9 (d, $^{2}J_{C-F}$ = 20.2 Hz, CH₂, C-7), 25.8 (s, CH₃, (CH₃)₃C), 21.0 (s, CH₃, CH₃CO), 18.3 (s, C, (CH₃)₃C), 16.3 (d, $^{3}J_{C-F}$ = 17.4 Hz, CH₂, C-6), -5.4 (s, CH₃, CH₃Si), -5.5 (s, CH₃, CH₃Si); 19 F NMR (235 MHz, CDCl₃) δ -144.1 to -144.4 (m). HRMS (ESI+) calcd for $[C_{15}H_{27}FO_4Si+Na]^+$ 341.1555, found 341.1550.

35: $[\alpha]_D$ –78.3 (c 0.60, CHCl₃); IR (ATR) 2929, 1748, 1229, 1100, 1010 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 6.18 (s, 1H, H-2), 4.05 (ddd, $J_{4,8}$ = 8.4 Hz, $J_{4,8}$ = 5.7 Hz, $J_{4,F}$ = 3.0 Hz, 1H, H-4), 3.71 (dd, $J_{\rm gem}$ = 10.2 Hz, $J_{8,4}$ = 5.7 Hz, 1H, H-8), 3.56 (dd, $J_{\rm gem}$ = 10.2 Hz, $J_{8,4}$ = 8.4 Hz, 1H, H-8), 3.01 (m, 1H, H-5), 2.49 (m, 1H, H-7exo), 2.36 (m, 1H,

H-7endo), 2.24 (m, 1H, H-6exo), 2.08 (s, 3H, CH₃CO), 1.50 (m, 1H, H-6endo), 0.88 (s, 9H, (CH₃)₃C), 0.07 (s, 3H, CH₃Si), 0.06 (s, 3H, CH₃Si); 13 C NMR (90 MHz, CDCl₃) δ 169.7 (s, C=O), 99.2 (d, $^{1}J_{C-F}$ = 243.7 Hz, C, C-1), 99.3 (d, $^{2}J_{C-F}$ = 18.2 Hz, CH, C-2), 87.4 (d, $^{3}J_{C-F}$ = 2.8 Hz, CH, C-4), 64.5 (s, CH₂, C-8), 44.2 (d, $^{2}J_{C-F}$ = 19.4 Hz, CH, C-5), 28.7 (d, $^{2}J_{C-F}$ = 23.9 Hz, CH₂, C-7), 25.9 (s, CH₃, (CH₃)₃C), 21.2 (s, CH₃, CH₃CO), 18.3 (s, C, (CH₃)₃C), 16.9 (d, $^{3}J_{C-F}$ = 12.7 Hz, CH₂, C-6), -5.3 (s, CH₃, CH₃Si), -5.4 (s, CH₃, CH₃Si); 19 F NMR (235 MHz, CDCl₃) δ -155.7 to -155.9 (m). HRMS (ESI+) calcd for [C₁₅H₂₇FO₄Si + Na] $^{+}$ 341.1555, found 341.1542.

(1S,4S,5R)-4-tert-Butyldimethylsilyloxymethyl-1-chloro-3-oxabicyclo[3.2.0]heptan-2-one (17a). To a solution of 17 (490 mg, 2.77 mmol) in CH₂Cl₂ (20 mL) at 0 °C were successively added imidazole (390 mg, 5.7 mmol) and tert-butyldimethylsilyl chloride (580 mg, 3.87 mmol). The mixture was allowed to warm to room temperature and stirred for 16 h. The reaction mixture was successively washed with 5% aqueous citric acid solution (3 \times 10 mL), saturated aqueous NaHCO₃ solution (3 \times 10 mL), and brine (3 \times 10 mL). Then, the organic layer was dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness affording 750 mg of a reaction crude which was used in the next reaction without further purification due to its instability. However, a small sample was purified allowing its complete characterization: $[\alpha]_D$ -6.7 (c 1.1, CHCl₃); IR (ATR) 2927, 2855, 1766, 834, 778 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 4.37 (t, $J_{4.8}$ = 4.6 Hz, 1H, H-4), 3.73 (d, $J_{8,4}$ = 4.6 Hz, 2H, H-8), 3.28 (dd, J = 8.9 Hz, J = 6.6 Hz, 1H, H-5), 2.71-2.45 (m, 3H, H-6, 2H-7), 1.83 (m, 1H, H-6), 0.89 (s, 9H, $(CH_3)_3C$), 0.07 (s, 6H, 2CH₃Si); ¹³C NMR (62.5 MHz, CDCl₃) δ 175.3 (C=O, C-2), 84.1 (CH, C-4), 63.6 (CH₂, C-8), 59.6 (C, C-1), 45.7 (CH, C-5), 33.8 (CH₂, C-7), 25.8 (CH₃, CH₃CO), 20.3 (CH₂, C-6), 18.4 (C, $(CH_3)_3C$), -3.7 (CH_3, CH_3Si) , -3.7 (s, CH_3, CH_3Si) . HRMS (ESI+)calcd for $[C_{13}H_{23}ClO_3Si + Na]^+$ 313.0997, found 313.0991.

(1S,2R,4S,5R)-2-Acetyloxy-4-tert-butyldimethylsilyloxymethyl-1-chloro-3-oxabicyclo[3.2.0] heptane (36). To a solution of crude 17a (750 mg, 2.57 mmol) in dry CH_2Cl_2 (10 mL) at -78 °C was added dropwise a 1.0 M solution of DIBAL-H in CH₂Cl₂ (4.9 mL, 4.9 mmol). After 3.5 h of stirring at -78 °C, the reaction mixture was quenched by the slow addition of a HNO₃ 3% solution (9 mL) and allowed to warm to room temperature. The two layers were separated, and the aqueous was extracted with CH_2Cl_2 (3 \times 10 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness affording a reaction crude which was used in the next reaction without further purification. To an ice-cooled solution of this crude and pyridine (2.8 mL, 36.1 mmol) in CH₂Cl₂ (25 mL) was added dropwise acetic anhydride (2.1 mL, 19.0 mmol). The reaction mixture was stirred at room temperature overnight. Then, the reaction was washed with HCl 3% (3 \times 15 mL), saturated aqueous NaHCO₃ solution (3 \times 15 mL), and brine (3 \times 15 mL). The organic layer was dried with anhydrous Na₂SO₄, filtered, and evaporated to dryness. The crude residue was purified by column chromatography (hexane-diethyl ether 10:1) to afford the α-acetate 36 (758 mg, 2.26 mmol, 82% yield over 3 steps) as a colorless oil: $[\alpha]_D$ +45.7 (c 1.40, CHCl₃); IR (ATR) 2929, 1751, 1221 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 6.37 (s, 1H, H-2), 4.15 (ddd, $J_{4,8}$ = 6.0 Hz, $J_{4,8}$ = 4.6 Hz, $J_{4,5}$ = 2.4 Hz, 1H, H-4), 3.69 (dd, $J_{\text{gem}} = 10.6$ Hz, $J_{8,4} = 4.6$ Hz, 1H, H-8), 3.62 (dd, $J_{\text{gem}} = 10.6 \text{ Hz}, J_{8,4} = 6.0 \text{ Hz}, 1\text{H}, \text{H-8}), 3.08 \text{ (m, 1H, H-5)}, 2.62 \text{ (m, 1H, H-6)}$ H-7endo), 2.37 (m, 2H, H-7exo, H-6exo), 2.15 (s, 3H, CH₃CO), 1.74 (m, 1H, H-6endo), 0.89 (s, 9H, (CH₃)₃C), 0.08 (s, 3H, CH₃Si), 0.07 (s, 3H, CH₃Si); 13 C NMR (62.5 MHz, CDCl₃) δ 169.5 (C=O), 103.8 (CH, C-2), 87.2 (CH, C-4), 71.1 (C, C-1), 64.4 (CH₂, C-8), 49.7 (CH, C-5), 30.2 (CH₂, C-7), 25.9 (CH₃, (CH₃)₃C), 21.1 (CH₃, CH₃CO), 20.9 (CH₂, C-6), 18.3 $(C, (CH_3)_3C), -5.4 (CH_3, CH_3Si), -5.4 (CH_3, CH_3Si). HRMS (ESI+)$ calcd for $[C_{15}H_{27}ClO_4Si + Na]^+$ 357.1259, found 357.1245.

(1'S,2'S,4'S,5'R)- and (1'S,2'R,4'S,5'R)-6-Chloro-9-(4'-tert-butyldimethylsilyloxymethyl-1'-fluoro-3'-oxabicyclo[3.2.0]hept-2-yl)-9H-purine (**37** and **38**) and (1'S,2'S,4'S,5'R)- and (1'S,2'R,4'S,5'R)-6-Chloro-7-(4'-tert-butyldimethylsilyloxymethyl-1'-fluoro-3'-oxabicyclo [3.2.0] hept-2-yl)-7H-purine (**39** and **40**). BSA (567 μ L, 2.10 mmol)

was added to a suspension of 6-chloropurine (172 mg, 1.10 mmol) in dry toluene (4 mL) under argon atmosphere. The reaction was stirred for 20 min and cooled to 0 °C. Then, a solution of a mixture of 34 and 35 (245 mg, 0.77 mmol) in dry toluene (2 mL) and TMSOTf (175 μ L, 0.95 mmol) was successively added, and the reaction mixture was stirred at 100 °C for 1 h. Then, after allowing the solution to cool to room temperature, CH₂Cl₂ (10 mL) was added, and the reaction was quenched with aqueous saturated NaHCO₃ (4 mL). The two layers were separated, and the aqueous layer was extracted with CH_2Cl_2 (3 \times 10 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness. The crude residue was purified by column chromatography (from hexane-EtOAc 10:1 to hexane-EtOAc 2:1) to afford the following fractions: (i) 37 (161 mg, 0.39 mmol, 53% yield) as a white solid; (ii) 38 (94 mg, 0.23 mmol, 30% yield) as a colorless oil; (iii) 39 (9 mg, 0.02 mmol, 3% yield) as a colorless oil; and (iv) 40 (9 mg, 0.02 mmol, 3% yield) as a yellow solid.

37: Mp decomposes over 60 °C (from hexane—EtOAc); $[\alpha]_D$ –13.8 (c 1.60, CHCl₃); IR (ATR) 3108, 2928, 1588, 1561, 1090 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.77 (s, 1H, H-2), 8.42 (s, 1H, H-8), 6.63 $(d, J_{2',F} = 15.6 \text{ Hz}, 1H, H-2'), 4.33 (ddd, J_{4',F} = 4.3 \text{ Hz}, J_{4',8'} = 4.1 \text{ Hz}, J_{4',8'} =$ 3.5 Hz, 1H, H-4'), 3.86 (dd, $J_{\text{gem}} = 10.9$ Hz, $J_{8',4'} = 4.1$ Hz, 1H, H-8'), 3.74 (dd, $J_{\text{gem}} = 10.9 \text{ Hz}$, $J_{8',4'} = 3.5 \text{ Hz}$, 1H, H-8'), 3.33 (m, 1H, H-5'), 2.29 (m, 2H, H-6'exo, H-7'exo), 1.59 (m, 1H, H-7'endo), 1.48 (m, 1H, H-6'endo), 0.93 (s, 9H, (CH₃)₃C), 0.11 (s, 3H, CH₃Si), 0.10 (s, 3H, CH₃Si); ¹³C NMR (62.5 MHz, CDCl₃) δ 152.4 (s, CH, C-2), 151.1 (2s, 2C, C-4/C-6), 142.6 (s, CH, C-8), 132.0 (s, C, C-5), 101.9 (d, ${}^{1}J_{C-F} = 238.5 \text{ Hz}$, C, C-1'), 91.6 (d, ${}^{2}J_{C-F}$ = 40.7 Hz, CH, C-2'), 86.1 (d, ${}^{3}J_{C-F}$ = 2.0 Hz, CH, C-4'), 65.1 (s, CH₂, C-8'), 45.9 (d, ${}^{2}J_{C-F}$ = 18.8 Hz, CH, C-5'), 25.8 (s, CH₃, $(CH_3)_3C$), 25.7 (d, ${}^2J_{C-F}$ = 24.6 Hz, CH_2 , C-7'), 18.3 (s, C, $(CH_3)_3C$), 16.1 (d, ${}^{3}J_{C-F}$ = 15.5 Hz, CH₂, C-6'), -5.5 (s, CH₃, 2CH₃Si); ${}^{19}F$ NMR (235 MHz, CDCl₃) δ -142.7 to -143.1 (m). HRMS (ESI+) calcd for $[C_{18}H_{26}CIFN_4O_2Si + Na]^+$ 435.1390, found 435.1379.

38: $[\alpha]_D - 16.0$ (c 0.50, CHCl₃); IR (ATR) 3128, 2929, 1702, 1590, 1097 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.73 (s, 1H, H-2), 8.54 (d, $J_{8,F}$ = 2.7 Hz, 1H, H-8), 6.58 (d, $J_{2',F}$ = 11.5 Hz, 1H, H-2'), 4.21 (ddd, $J_{4',F}$ = 5.6 Hz, $J_{4',8'}$ = 3.6 Hz, $J_{4',8'}$ = 3.6 Hz, 1H, H-4'), 3.87 (dd, J_{gem} = 11.2 Hz, $J_{8',4'}$ = 3.6 Hz, 1H, H-8'), 3.28 (m, 1H, H-8'), 3.74 (dd, J_{gem} = 11.2 Hz, $J_{8',4'}$ = 3.6 Hz, 1H, H-8'), 2.58 (m, 1H, H-7'endo), 2.50 (m, 1H, H-7'exo), 2.36 (m, 1H, H-6'exo), 1.62 (m, 1H, H-6'endo), 0.93 (s, 9H, (CH₃)₃C), 0.13 (s, 3H, CH₃Si), 0.11 (s, 3H, CH₃Si); ¹³C NMR (62.5 MHz, CDCl₃) δ 152.0 (s, CH, C-2), 151.6/150.9 (2s, 2C, C-4/C-6), 144.9 (d, $^4J_{C-F}$ = 4.6 Hz, CH, C-8), 131.4 (s, C, C-5), 98.0 (d, $^1J_{C-F}$ = 250.6 Hz, C, C-1'), 89.3 (d, $^2J_{C-F}$ = 17.8 Hz, CH, C-2'), 85.8 (s, CH, C-4'), 64.5 (s, CH₂, C-8'), 47.2 (d, $^2J_{C-F}$ = 18.7 Hz, CH, C-5'), 28.6 (d, $^2J_{C-F}$ = 22.8 Hz, CH₂, C-7'), 25.9 (s, CH₃, (CH₃)₃C), 18.4 (s, C, (CH₃)₃C), 16.6 (d, $^3J_{C-F}$ = 18.2 Hz, CH₂, C-6'), -5.4 (s, CH₃, 2CH₃Si); ¹⁹F NMR (235 MHz, CDCl₃) δ -145.7 to -146.0 (m).HRMS (ESI+) calcd for $[C_{18}H_{26}CIFN_4O_2Si+Na]^+$ 435.1390, found 435.1377.

39: Mp 61–63 °C (from EtOAc); $[\alpha]_D$ –63.1 (c 1.60, CHCl₃); IR (ATR) 2927, 1377, 1254, 1099 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 8.89 (s, 1H, H-2), 8.69 (s, 1H, H-8), 6.98 (d, $J_{2',F}$ = 14.0 Hz, 1H, H-2'), 4.39 (ddd, $J_{4',8'}$ = 3.5 Hz, $J_{4',F}$ = 3.3 Hz, $J_{4',8'}$ = 3.0 Hz, 1H, H-4'), 3.90 (dd, $J_{\text{gem}} = 11.0 \text{ Hz}$, $J_{8',4'} = 3.5 \text{ Hz}$, 1H, H-8'), 3.73 (dd, $J_{\text{gem}} = 11.0 \text{ Hz}$, $J_{8',4'} = 3.0$ Hz, 1H, H-8'), 3.23 (m, 1H, H-5'), 2.28 (m, 2H, H-6'exo, H-7'exo), 1.47 (m, 1H, H-7'endo), 1.39 (m, 1H, H-6'endo), 0.92 (s, 9H, (CH₃)₃C), 0.11 (s, 3H, CH₃Si), 0.10 (s, 3H, CH₃Si); 13 C NMR (90 MHz, CDCl₃) δ 162.2 (s, C, C-4), 152.6 (s, CH, C-2), 146.1 (s, CH, C-8), 143.2 (s, C, C-6), 121.8 (s, C, C-5), 101.9 (d, ${}^{1}J_{C-F}$ = 240.0 Hz, C, C-1'), 93.1 (d, ${}^{2}J_{C-F}$ = 38.7 Hz, CH, C-2'), 86.4 (d, ${}^{3}J_{C-F} = 1.7$ Hz, CH, C-4'), 65.5 (s, CH₂, C-8'), 46.0 (d, ${}^{2}J_{C-F}$ = 19.0 Hz, CH, C-5'), 25.8 (s, CH₃, (CH₃)₃C), 25.5 (d, ${}^{2}J_{C-F}$ = 23.3 Hz, CH₂, C-7'), 18.3 (s, C, (CH₃)₃C), 16.4 (d, ${}^{3}J_{C-F}$ = 15.6 Hz, CH₂, C-6'), -5.5 (s, CH₃, 2CH₃Si); ¹⁹F NMR (235 MHz, CDCl₃) δ -142.5to -142.6 (m). HRMS (ESI+) calcd for $[C_{18}H_{26}ClFN_4O_2Si + Na]^+$ 435.1390, found 435.1374.

40: $[\alpha]_D$ +41.8 (c 1.10, CHCl₃); IR (ATR) 2928, 1599, 1536, 1257, 838 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.89 (d, $J_{8,F}$ = 1.6 Hz, 1H, H-8), 8.87 (d, $J_{2,F}$ = 1.2 Hz, 1H, H-2), 6.80 (d, $J_{2',F}$ = 8.2 Hz, 1H, H-2'), 4.27 (ddd, $J_{4',F}$ = 5.5 Hz, $J_{4',8'}$ = 4.2 Hz, $J_{4',8'}$ = 3.2 Hz, 1H, H-4'), 3.92 (dd, $J_{\text{gem}} = 11.5 \text{ Hz}$, $J_{8',4'} = 3.2 \text{ Hz}$, 1H, H-8'), 3.72 (dd, $J_{\text{gem}} = 11.5 \text{ Hz}$, $J_{8',4'} = 4.2 \text{ Hz}$, 1H, H-8'), 3.22 (m, 1H, H-5'), 2.68 (m, 1H, H-7'endo), 2.59 (m, 1H, H-7'exo), 2.36 (m, 1H, H-6'exo), 1.62 (m, 1H, H-6'endo), 0.91 (s, 9H, $(CH_3)_3C$), 0.13 (s, 3H, CH_3Si), 0.09 (s, 3H, CH_3Si); ¹³C NMR (90 MHz, CDCl₃) δ 162.4 (s, C, C-4), 152.2 (s, CH, C-2), 148.4 (d, ${}^{4}J_{C-F}$ = 2.4 Hz, CH, C-8), 141.7 (s, C, C-6), 121.9 (s, C, C-5), 99.1 $(d, {}^{1}J_{C-F} = 249.7 \text{ Hz}, C, C-1'), 91.7 (d, {}^{2}J_{C-F} = 18.1 \text{ Hz}, CH, C-2'), 86.5$ (s, CH, C-4'), 64.3 (s, CH₂, C-8'), 46.7 (d, ${}^{2}J_{C-F}$ = 19.1 Hz, CH, C-5'), 29.2 (d, ${}^{2}J_{C-F}$ = 23.0 Hz, CH₂, C-7'), 25.8 (s, CH₃, (CH₃)₃C), 18.3 (s, C, $(CH_3)_3C$), 16.3 (d, ${}^3J_{C-F} = 16.7 \text{ Hz}$, CH_2 , C-6'), -5.5 (s, CH_3 , 2CH₃Si); ¹⁹F NMR (235 MHz, CDCl₃) δ –146.1 to –146.3 (m). HRMS (ESI+) calcd for $[C_{18}H_{26}ClFN_4O_2Si + Na]^+$ 435.1390, found 435.1383.

(1'S,2'S,4'S,5'R)- and (1'S,2'R,4'S,5'R)-6-Chloro-9-(4'-tert-butvldimethylsilyloxymethyl-1'-chloro-3'-oxabicyclo[3.2.0]hept-2-yl)-9Hpurine (41 and 42) and (1'S,2'S,4'S,5'R)- and (1'S,2'R,4'S,5'R)-6-Chloro-7-(4'-tert-butyldimethylsilyloxymethyl-1'-chloro-3'-oxabicyclo [3.2.0]hept-2-yl)-7H-purine (43 and 44). BSA (147 μ L, 0.57 mmol) was added to a suspension of 6-chloropurine (44 mg, 0.29 mmol) in dry acetonitrile (2 mL) under an argon atmosphere. The reaction was stirred for 20 min and cooled to 0 °C. Then, a solution of 36 (63 mg, 0.19 mmol) in dry acetonitrile (1.5 mL) and TMSOTf (45 μ L, 0.25 mmol) was successively added, and the reaction mixture was stirred at 82 °C for 1 h. Then CH₂Cl₂ (5 mL) was added, and the reaction was guenched with aqueous saturated NaHCO₃ (3 mL), after allowing the solution to cool to room temperature. The mixture was separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 5 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness. The crude residue was purified by column chromatography (from hexane-EtAcO 14:1 to hexane-EtAcO 1:1) to afford the following fractions: (i) 42 (10 mg, 0.02 mmol, 12% yield) as a colorless oil; (ii) 41 (57 mg, 0.13 mmol, 70% yield) as a white solid; (iii) 43 (4 mg, 0.01 mmol, 5% yield) as a colorless oil; and (iv) 44 (6 mg, 0.02 mmol, 8% yield) as a colorless oil.

41: Mp decomposes over 70 °C (from hexane—EtAcO); $[\alpha]_D + 17.2$ (c 2.5, CHCl₃); IR (ATR) 3112, 2949, 2854, 1590, 1561, 1099 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.79 (s, 1H, H-2), 8.45 (s, 1H, H-8), 6.72 (s, 1H, H-2'), 4.34 (ddd, $J_{4',8'}$ = 4.7 Hz, $J_{4',8'}$ = 4.1 Hz, $J_{4',5'}$ = 1.0 Hz, 1H, H-4'), 3.85 (dd, J_{gem} = 10.9 Hz, $J_{8',4'}$ = 4.7 Hz, 1H, H-8'), 3.77 (dd, J_{gem} = 10.9 Hz, $J_{8',4'}$ = 4.1 Hz, 1H, H-8'), 3.29 (ddd, $J_{5',6'}$ = 8.2 Hz, $J_{5',6'}$ = 5.8 Hz, $J_{5',4'}$ = 1.0 Hz, 1H, H-5'), 2.55 (m, 1H, H-6'exo), 2.33 (m, 1H, H-7'exo), 2.12 (m, 1H, H-7'endo), 1.82 (m, 1H, H-6'endo), 0.94 (s, 9H, (CH₃)₃C), 0.13 (s, 3H, CH₃Si), 0.12 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 152.4 (CH, C-2), 151.4/151.1 (2C, C-4/C-6), 142.7 (CH, C-8), 131.8 (C, C-5), 93.8 (CH, C-2'), 86.3 (CH, C-4'), 70.5 (C, C-1'), 64.7 (CH₂, C-8'), 49.7 (CH, C-5'), 29.7 (CH₂, C-7'), 25.9 (CH₃, (CH₃)₃C), 20.2 (CH₂, C-6'), 18.4 (C, (CH₃)₃C), -5.4 (s, CH₃, 2CH₃Si). HRMS (ESI+) calcd for $[C_{18}H_{26}Cl_2N_4O_2Si+Na]^+$ 451.1094, found 451.1094.

42: $[\alpha]_D$ – 30.0 (c 0.90, CHCl₃); IR (ATR) 2928, 2856, 1589, 1561, 1215 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.71 (s, 1H, H-2), 8.50 (s, 1H, H-8), 6.63 (s, 1H, H-2'), 4.25 (ddd, $J_{4',8'}$ = 4.5 Hz, $J_{4',8'}$ = 4.5 Hz, $J_{4',8'}$ = 4.5 Hz, 1H, H-8'), 3.80 (dd, J_{gem} = 11.0 Hz, $J_{8',4'}$ = 4.5 Hz, 1H, H-8'), 3.31 (m, 1H, H-5'), 2.91 (m, 1H, H-7'endo), 2.61 (m, 2H, H-6'exo, H-7'exo), 1.97 (m, 1H, H-6'endo), 0.93 (s, 9H, (CH₃)₃C), 0.13 (s, 3H, CH₃Si), 0.12 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 152.1 (CH, C-2), 151.2/150.8 (2C, C-4/C-6), 144.0 (CH, C-8), 131.6 (C, C-5), 91.9 (CH, C-2'), 86.4 (CH, C-4'), 71.7 (C, C-1'), 61.6 (CH₂, C-8'), 50.8 (CH, C-5'), 33.8 (CH₂, C-7'), 25.9 (CH₃, (CH₃)₃C), 21.5 (CH₂, C-6'), 18.4 (C, (CH₃)₃C), -5.3 (s, CH₃,

CH₃Si), -5.4 (s, CH₃, CH₃Si). HRMS (ESI+) calcd for [C₁₈H₂₆Cl₂-N₄O₂Si + Na]⁺ 451.1094, found 451.1083.

43: $[\alpha]_D$ - 36.8 (c 1.25, CHCl₃); IR (ATR) 2928, 2856, 1680, 1094 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.91 (s, 1H, H-2), 8.68 (s, 1H, H-8), 7.00 (s, 1H, H-2'), 4.37 (ddd, $J_{4',8'}$ = 3.9 Hz, $J_{4',8'}$ = 3.6 Hz, $J_{4',5'}$ = 1.1 Hz, 1H, H-4'), 3.90 (dd, J_{gem} = 11.0 Hz, $J_{8',4'}$ = 3.9 Hz, 1H, H-8'), 3.76 (dd, J_{gem} = 11.0 Hz, $J_{8',4'}$ = 3.6 Hz, 1H, H-8'), 3.30 (ddd, $J_{5',6'}$ = 8.2 Hz, $J_{5',6'}$ = 5.8 Hz, $J_{5',4'}$ = 1.1 Hz, 1H, H-5'), 2.58 (m, 1H, H-6'exo), 2.36 (m, 1H, H-7'exo), 2.12 (m, 1H, H-7'endo), 1.78 (m, 1H, H-6'endo), 0.93 (s, 9H, (CH₃)₃C), 0.12 (s, 3H, CH₃Si), 0.12 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 162.1 (C, C-4), 152.7 (CH, C-2), 146.4 (CH, C-8), 143.1 (C, C-6), 122.1 (C, C-5), 94.3 (CH, C-2'), 86.6 (CH, C-4'), 70.8 (C, C-1'), 65.3 (CH₂, C-8'), 49.8 (CH, C-5'), 30.1 (CH₂, C-7'), 26.0 (CH₃, (CH₃)₃C), 20.7 (CH₂, C-6'), 18.4 (C, (CH₃)₃C), -5.4 (CH₃, 2CH₃Si). HRMS (ESI+) calcd for $[C_{18}H_{26}Cl_2N_4O_2Si + Na]^+$ 451.1094, found 451.1091.

44: $[\alpha]_D$ +52.5 (c 0.40, CHCl₃); IR (ATR) 2928, 2856, 1252, 1052 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.91 (s, 1H, H-8), 8.89 (s, 1H, H-2), 6.90 (s, 1H, H-2'), 4.31 (ddd, $J_{4',8'}$ = 4.6 Hz, $J_{4',8'}$ = 3.7 Hz, $J_{4',5'}$ = 3.1 Hz, 1H, H-4'), 3.94 (dd, $J_{\rm gem}$ = 11.3 Hz, $J_{8',4'}$ = 3.7 Hz, 1H, H-8'), 3.80 (dd, $J_{\rm gem}$ = 11.3 Hz, $J_{8',4'}$ = 4.6 Hz, 1H, H-8'), 3.28 (m, 1H, H-5'), 2.98 (m, 1H, H-7'endo), 2.73 (m, 1H, H-7'exo), 2.59 (m, 1H, H-6'exo), 1.92 (m, 1H, H-6'endo), 0.90 (s, 9H, (CH₃)₃C), 0.15 (s, 3H, CH₃Si), 0.14 (s, 3H, CH₃Si); ¹³C NMR (90 MHz, CDCl₃) δ 162.4 (C, C-4), 152.4 (CH, C-2), 147.7 (CH, C-8), 142.0 (C, C-6), 122.0 (C, C-5), 93.9 (CH, C-2'), 87.1 (CH, C-4'), 72.9 (C, C-1'), 63.8 (CH₂, C-8'), 50.7 (CH, C-5'), 34.5 (CH₂, C-7'), 26.0 (CH₃, (CH₃)₃C), 20.6 (CH₂, C-6'), 18.4 (C, (CH₃)₃C), -5.3 (CH₃, 2CH₃Si). HRMS (ESI+) calcd for $[C_{18}H_{26}Cl_2N_4O_2Si + Na]^+$ 451.1094, found 451.1089.

(1'S,2'S,4'S,5'R)-6-Chloro-9-(1'-fluoro-4'-hydroxymethyl-3'-oxabicyclo [3.2.0]hept-2-yl)-9H-purine (45). To an ice-cooled solution of 37 (80 mg, 0.19 mmol) in THF (5 mL) was added Et₃N \cdot 3HF (47 μ L, 0.29 mmol), and the resulting solution was stirred for 5 h at room temperature. Then, more Et₃N·3HF (47 μ L, 0.29 mmol) was added, and after 24 h of stirring at room temperature, the solvent was removed. The residue was purified by column chromatography (EtOAc-hexane 2:1) to afford 45 (52 mg, 0.17 mmol, 90% yield) as a white solid: mp 123–126 °C (from CHCl₃); $[\alpha]_D$ –13.3 (c 1.05, CHCl₃); IR (ATR) 3373, 2938, 1590, 1562, 1206, 1034 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.78 (s, 1H, H-2), 8.44 (s, 1H, H-8), 6.64 (dd, $J_{2',F} = 15.9 \text{ Hz}, J = 0.9 \text{ Hz}, 1\text{H}, \text{H}-2'), 4.45 \text{ (ddd}, J = 5.1 \text{ Hz}, J = 5.0 \text{ Hz}, J = 4.8$ Hz, 1H, H-4'), 3.83 (m, 2H, H-8'), 3.19 (m, 1H, H-5'), 2.32 (m, 2H, H-6'exo, H-7'exo), 2.16 (t, $J_{OH,8'} = 5.3$ Hz, 1H, OH), 1.61 (m, 1H, H-7'endo), 1.53 (m, 1H, H-6'endo); 13 C NMR (90 MHz, CDCl₃) δ 152.5 (s, CH, C-2), 151.3/150.9 (2s, 2C, C-4/C-6), 142.5 (s, CH, C-8), 131.9 (s, C, C-5), 103.2, 101.9 (d, ${}^{1}J_{C-F}$ = 283.2 Hz, C, C-1'), 91.2 (d, ${}^{2}J_{C-F}$ = 41.2 Hz, CH, C-2'), 86.1 (d, ${}^{3}J_{C-F} = 1.3$ Hz, CH, C-4'), 63.8 (s, CH₂, C-8'), 45.6 $(d, {}^{2}J_{C-F} = 18.7 \text{ Hz}, CH, C-5'), 25.6 (d, {}^{2}J_{C-F} = 23.2 \text{ Hz}, CH_{2}, C-7'), 15.9$ (d, ${}^{3}J_{C-F}$ = 15.7 Hz, CH₂, C-6'); ${}^{19}F$ NMR (235 MHz, CDCl₃) δ –142.1 to -142.4 (m). HRMS (ESI+) calcd for $[C_{12}H_{12}CIFN_4O_2 + Na]^+$ 321.0525, found 321.0530.

(1'S,2'R,4'S,5'R)-6-Chloro-9-(1'-fluoro-4'-hydroxymethyl-3'-oxabicyclo [3.2.0]hept-2-yl)-9H-purine (**46**). To an ice-cooled solution of **38** (54 mg, 0.13 mmol) in THF (3 mL) was added Et₃N · 3HF (65 μL, 0.39 mmol), and the resulting solution was stirred for 7 h at room temperature. Then, more Et₃N · 3HF (65 μL, 0.39 mmol) was added, and after 24 h of stirring at room temperature, the solvent was removed. The residue was purified by column chromatography (EtOAc—hexane 2:1) to give **46** (35 mg, 0.12 mmol, 90% yield) as a white solid: mp 283–287 °C (from CHCl₃); [α]_D −16.7 (c 0.90, MeOH); IR (ATR) 3313, 2924, 1592, 1562, 1204, 1141 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.75 (s, 1H, H-2), 8.41 (d, $J_{8,F}$ = 2.0 Hz, 1H, H-8), 6.47 (d, $J_{2',F}$ = 12.2 Hz, 1H, H-2'), 4.30 (ddd, $J_{4',F}$ = 5.7 Hz, $J_{4',8'}$ = 4.4 Hz, $J_{4',5'}$ = 2.4 Hz, 1H, H-4'), 3.96 (dd, J_{gem} = 12.2 Hz, $J_{8',4'}$ = 2.4 Hz, 1H, H-8'), 3.74 (dd, J_{gem} = 12.2 Hz, $J_{8',4'}$ = 4.4 Hz, 1H, H-8'), 3.41 (m, 1H, H-5'), 2.56 (m, 2H, H-7' exo, H-7' endo), 2.41 (m, 1H, H-6' exo), 1.63 (m,

1H, H-6' endo); 13 C NMR (90 MHz, CDCl₃) δ 151.9 (s, CH, C-2), 151.6/151.5 (2s, 2C, C-4/C-6), 144.6 (d, $^4J_{C-F}$ = 3.3 Hz, CH, C-8), 132.0 (s, C, C-5), 98.4 (d, $^1J_{C-F}$ = 250.6 Hz, C, C-1'), 90.9 (d, $^2J_{C-F}$ = 18.1 Hz, CH, C-2'), 86.3 (s, CH, C-4'), 63.9 (s, CH₂, C-8'), 46.8 (d, $^2J_{C-F}$ = 18.6 Hz, CH, C-5'), 28.6 (d, $^2J_{C-F}$ = 22.9 Hz, CH₂, C-7'), 16.5 (d, $^3J_{C-F}$ = 17.8 Hz, CH₂, C-6'); 19 F NMR (235 MHz, CDCl₃) δ —146.0 to —146.2 (m). HRMS (ESI+) calcd for [C₁₂H₁₂CIFN₄O₂ + Na]⁺ 321.0525, found 321.0522. Anal. Calcd for (C₁₂H₁₂CIFN₄O₂): C, 48.25; H, 4.05; N, 18.76. Found: C, 48.12; H, 4.18; N, 18.62.

(1'S,2'S,4'S,5'R)-9-(1'-Fluoro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-adenine (49). A solution of 45 (20 mg, 0.07 mmol) and saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 40 h. After cooling at room temperature, the solvent was removed under vacuum, and the residue was purified by column chromatography (CH₂Cl₂-MeOH 10:1) to furnish 49 (17 mg, 0.06 mmol, 89% yield) as a white solid: mp 210–213 °C (from MeOH); $[\alpha]_D$ –45.7 (c 0.35, MeOH); IR (ATR) 3326, 3214, 2920, 2852, 1651, 1600, 1080, 1015 cm⁻¹; ¹H NMR (360 MHz, DMSO-d₆) δ 8.41 (s, 1H, H-8), 8.14 (s, 1H, H-2), 7.34 (br s, 2H, NH₂), 6.48 (d, $J_{2',F}$ = 16.7 Hz, 1H, H-2'), 5.05 (t, $J_{OH,8'}$ = 5.6 Hz, $J_{OH,8'}$ = 5.0 Hz, 1H, OH), 4.27 (t, J = 5.0 Hz, 1H, H-4'), 3.54 (m, 2H, H-8'), 3.57 (m, 1H, H-5'), 2.18 (m, 2H, H-6'exo, H-7'exo), 1.59 (m, 2H, H-6'endo, H-7'endo); ¹³C NMR (90 MHz, DMSO- d_6) δ 155.9 (s, C, C-6), 152.8 (s, CH, C-2), 148.9 (s, C, C-4), 138.1 (s, CH, C-8), 118.7 (s, C, C-5), 102.0 (d, ${}^{1}J_{C-F}$ = 235.4 Hz, C, C-1'), 89.8 (d, ${}^2J_{C-F}$ = 40.8 Hz, CH, C-2'), 85.1 (s, CH, C-4'), 62.6 (s, CH₂, C-8'), 45.2 (d, ${}^2J_{C-F}$ = 18.1 Hz, CH, C-5'), 25.2 (d, ${}^2J_{C-F}$ = 23.0 Hz, CH₂, C-7'), 14.9 (d, ${}^3J_{C-F}$ = 16.5 Hz, CH₂, C-6'); ${}^{19}F$ NMR (235 MHz, DMSO- d_6) δ -140.3 to -140.6 (m). HRMS (ESI+) calcd for $[C_{12}H_{14}FN_5O_2 + H]^+$ 280.1204, found 280.1202.

(1'S,2'R,4'S,5'R)-9-(1'-Fluoro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-adenine (50). A solution of 46 (26 mg, 0.06 mmol) and saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 40 h. After cooling at room temperature, the solvent was removed under vacuum, and the residue was purified by column chromatography (CH₂Cl₂-MeOH, 13:1) to afford **50** (18 mg, 0.06 mmol, 75% yield) as a white solid: mp 248–251 °C (from MeOH); $[\alpha]_D$ –5.7 (c 0.35, MeOH); IR (ATR) 3328, 3104, 2926, 1726, 1606, 1300, 1081, 1014 cm $^{-1}$; ¹H NMR (360 MHz, DMSO- d_6) δ 8.28 (d, $J_{8,F}$ = 2.4 Hz, 1H, H-8), 8.15 (s, 1H, H-2), 7.32 (br s, 2H, NH₂), 6.50 (d, $J_{2',F}$ = 12.2 Hz, 1H, H-2'), 5.26 (t, $J_{OH,8'}$ = 5.1 Hz, $J_{OH,8'}$ = 4.8 Hz, 1H, OH), 4.16 (ddd, $J_{4',8'} = 5.4 \text{ Hz}, J_{4',8'} = 4.2 \text{ Hz}, J_{4',F} = 2.2 \text{ Hz}, 1\text{H}, \text{H-4'}), 3.61 \text{ (ddd, } J_{gem} = 1.0 \text$ 11.7 Hz, $J_{8',OH}$ = 4.8 Hz, $J_{8',4'}$ = 4.2 Hz, 1H, H-8'), 3.52 (ddd, J_{gem} = 11.7 Hz, $J_{8',4'} = 5.4$ Hz, $J_{8',OH} = 5.1$ Hz, 1H, H-8'), 3.24 (m, 1H, H-5'), 2.45 (m, 1H, H-7'exo), 2.34 (m, 1H, H-7'endo), 2.23 (m, 1H, H-6'exo), 1.62 (m, 1H, H-6'endo); 13 C NMR (90 MHz, DMSO- d_6) δ 155.9 (s, C, C-6), 152.6 (s, CH, C-2), 149.1 (s, C, C-4), 139.4 (d, ${}^{4}J_{C-F} = 3.4 \text{ Hz}$, CH, C-8), 118.2 (s, C, C-5), 98.1 (d, ${}^{1}J_{C-F}$ = 247.6 Hz, C, C-1'), 88.4 (d, ${}^{2}J_{C-F}$ = 17.7 Hz, CH, C-2'), 85.1 (s, CH, C-4'), 62.6 (s, CH₂, C-8'), 46.5 (d, $^2 J_{\rm C-F}$ = 18.3 Hz, CH, C-5′), 28.0 (d, $^2 J_{\rm C-F}$ = 22.5 Hz, CH₂, C-7′), 15.8 (d, $^3 J_{\rm C-F}$ = 17.9 Hz, CH₂, C-6′); $^{19} {\rm F}$ NMR (235 MHz, DMSO- $^4 d_0$) δ -146.2 to -146.5 (m). HRMS (ESI+) calcd for $[C_{12}H_{14}FN_5O_2 + H]^+$ 280.1204, found 280.1203.

(1'5,2'5,4'5,5'R)-9-(1'-Fluoro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-hypoxanthine (**53**). To a solution of **45** (34 mg, 0.11 mmol) and MeONa (26 mg, 0.45 mmol) in dioxane (2 mL) were successively added 2-mercaptoethanol (32 μL, 0.45 mmol) and 2 drops of H₂O at room temperature. The solution was heated at 110 °C for 15 h when the same initial amounts of MeONa, 2-mercaptoethanol, and H₂O were added, and heating was continued for 4 days. Then, the solution was neutralized with glacial AcOH. The solvent was removed under vacuum, and the residue was purified by column chromatography (EtOAc–MeOH 9:1) to give **53** (22 mg, 0.08 mmol, 69% yield) as a white solid: mp 244–246 °C (MeOH); [α]_D –20.0 (c 0.30, DMSO); IR (ATR) 3319, 3061, 2853, 1691, 1169, 1033 cm⁻¹; ¹H NMR (360 MHz, DMSO- d_6)

 δ 12.40 (br s, 1H, N—H), 8.37 (s, 1H, H-8), 8.10 (s, 1H, H-2), 6.46 (d, $J_{2',F}$ = 16.1 Hz, 1H, H-2′), 5.05 (br s, 1H, OH), 4.28 (dd, J = 4.6 Hz, J = 4.4 Hz, 1H, H-4′), 3.52 (m, 2H, H-8′), 3.17 (m, 1H, H-5′), 2.16 (m, 2H, H-6′exo/7′exo), 1.56 (m, 1H, H-6′endo/7′endo); $^{13}\mathrm{C}$ NMR (90 MHz, DMSO- d_6) δ 156.5 (s, C, C-6), 147.7 (s, C, C-4), 146.2 (s, CH, C-2), 137.5 (s, CH, C-8), 124.2 (s, C, C-5), 102.1 (d, $^1J_{C-F}$ = 235.9 Hz, C, C-1′), 90.2 (d, $^2J_{C-F}$ = 40.8 Hz, CH, C-2′), 85.4 (d, $^3J_{C-F}$ = 1.4 Hz, CH, C-4′), 62.6 (s, CH₂, C-8′), 45.2 (d, $^2J_{C-F}$ = 18.2 Hz, CH, C-5′), 25.2 (d, $^2J_{C-F}$ = 22.8 Hz, CH₂, C-7′), 14.9 (d, $^3J_{C-F}$ = 16.1 Hz, CH₂, C-6′); $^{19}\mathrm{F}$ NMR (235 MHz, DMSO- d_6) δ —142.2 to —142.5 (m). HRMS (ESI+) calcd for [C₁₂H₁₃FN₄O₃ + Na] $^+$ 303.0864, found 303.0860.

(1'S,2'R,4'S,5'R)-9-(1'-Fluoro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-hypoxanthine (54). To a solution of 46 (15 mg, 0.05 mmol) and MeONa (10 mg, 0.20 mmol) in dioxane (3 mL) were successively added 2-mercaptoethanol (11 μ L, 0.20 mmol) and 3 drops of H₂O at room temperature. The solution was heated at 110 °C for 22 h and then, after being cooled to room temperature, was neutralized with glacial AcOH. The solvent was removed under vacuum, and the residue was purified by column chromatography (EtOAc-MeOH 10:1) to give 54 (12 mg, 0.04 mmol, 86% yield) as a white solid: mp 127-129 °C (MeOH); $[\alpha]_D$ +8.0 (c 0.50, MeOH); IR (ATR) 3338, 3048, 2868, 1698, 1207, 1027; ¹H NMR (360 MHz, DMSO- d_6) δ 12.30 (br s, 1H, N-H), 8.23 (d, $J_{8,F}$ = 2.4 Hz, 1H, H-8), 8.08 (s, 1H, H-2), 6.43 (d, $J_{2',F}$ = 11.4 Hz, 1H, H-2'), 5.10 (br s, 1H, OH), 4.14 (ddd, $J_{4',5'} = 6.6$ Hz, $J_{4',8'} =$ 4.6 Hz, $J_{4',8'}$ = 4.2 Hz, 1H, H-4'), 3.59 (dd, J_{gem} = 11.7 Hz, $J_{8',4'}$ = 4.2 Hz, 1H, H-8'), 3.52 (dd, $J_{\text{gem}} = 11.7 \text{ Hz}$, $J_{8',4'} = 4.6 \text{ Hz}$, 1H, H-8'), 3.23 (m, 1H, H-5'), 2.41 (m, 2H, H-7'exo/7'endo), 2.23 (m, 1H, H-6'exo), 1.63 (m, 1H, H-6'endo); 13 C NMR (90 MHz, DMSO- d_6) δ 156.6 (s, C, C-6), 147.8 (s, C, C-4), 146.2 (s, CH, C-2), 138.6 (d, ${}^{4}J_{C-F}$ = 3.2 Hz, CH, C-8), 123.6 (s, C, C-5), 98.2 (d, ${}^{1}J_{C-F} = 247.7$ Hz, C, C-1'), 88.6 (d, ${}^{2}J_{C-F} =$ 17.7 Hz, CH, C-2'), 85.3 (s, CH, C-4'), 62.5 (s, CH₂, C-8'), 46.4 (d, $^{2}J_{\rm C-F}$ = 18.3 Hz, CH, C-5′), 28.1 (d, $^{2}J_{\rm C-F}$ = 22.4 Hz, CH₂, C-7′), 15.7 (d, $^{3}J_{\rm C-F}$ = 17.7 Hz, CH₂, C-6′); ¹⁹F NMR (235 MHz, DMSO- $^{4}d_{\rm 0}$) δ -140.4 to -140.6 (m). HRMS (ESI+) calcd for $[C_{12}H_{13}FN_4O_3 +$ Na]⁺ 303.0864, found 303.0856.

⁻ (1'S,2'S,4'S,5'R)-6-Chloro-9-(1'-chloro-4'-hydroxymethyl-3'-oxabicyclo [3.2.0]hept-2-yl)-9H-purine (47). To an ice-cooled solution of 41 (132 mg, 0.31 mmol) in THF (4 mL) was added a 1.0 M solution of TBAF in THF (460 μ L, 0.46 mmol), and the resulting solution was stirred for 1 h at room temperature. After removal of the solvent, the residue was purified by column chromatography (EtOAc-hexane 3:1) to afford 47 (85 mg, 0.27 mmol, 88% yield) as a white solid: mp 127–129 °C (from CHCl₃); $[\alpha]_D$ +29.7 (c 0.74, CHCl₃); IR (ATR) 3380, 3124, 2946, 1593, 1562, 1199 cm⁻¹; ¹H NMR (360 MHz, CDCl₃) δ 8.75 (s, 1H, H-2), 8.45 (s, 1H, H-8), 6.66 (d, J =0.6 Hz, 1H, H-2'), 4.43 (ddd, $J_{4',8'} = 6.5$ Hz, $J_{4',8'} = 4.0$ Hz, $J_{4',5'} = 1.0$ Hz, 1H, H-4'), 3.86 (ddd, $J_{gem} = 11.9 \text{ Hz}$, $J_{8',OH} = 6.3 \text{ Hz}$, $J_{8',4'} = 4.0 \text{ Hz}$, 1H, H-8'), 3.79 (ddd, $J_{\text{gem}} = 11.9 \text{ Hz}$, $J_{8',4'} = 6.5 \text{ Hz}$, $J_{8',\text{OH}} = 4.5 \text{ Hz}$, 1H, H-8'), 3.21 $(ddd, J_{5',6'} = 8.2 \text{ Hz}, J_{5',6'} = 6.0 \text{ Hz}, J_{5',4'} = 1.0 \text{ Hz}, 1\text{H}, \text{H}-5'), 2.91 (dd, J_{OH,6'})$ = 6.3 Hz, $J_{OH,8'}$ = 4.5 Hz, 1H, OH), 2.53 (m, 1H, H-6'exo), 2.35 (m, 1H, H-7'exo), 2.04 (m, 1H, H-7'endo), 1.82 (m, 1H, H-6'endo); 13C NMR (90 MHz, CDCl₃) δ 152.4 (CH, C-2), 151.3/151.2 (2C, C-4/C-6), 142.7 (CH, C-8), 131.7 (C, C-5), 93.4 (CH, C-2'), 86.4 (CH, C-4'), 70.2 (C, C-1'), 63.2 (CH₂, C-8'), 49.7 (CH, C-5'), 29.7 (CH₂, C-7'), 20.0 (CH₂, C-6'). HRMS (ESI+) calcd for $[C_{12}H_{12}Cl_2N_4O_2 + Na]^+$ 337.0230, found 337.0222.

(1'S,2'S,4'S,5'R)-9-(1'-Chloro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-adenine (**51**). A solution of 47 (25 mg, 0.08 mmol) in saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 68 h. After cooling at room temperature, the solvent was removed under vacuum, and the resulting residue was purified by column chromatography (CH₂Cl₂-MeOH, 10:1) to afford **51** (21 mg, 0.07 mmol, 91% yield) as a white solid: mp 90–93 °C (from MeOH); [α]_D +26.7 (α 0.60, MeOH); IR (ATR) 3327, 3168, 2947, 2852, 1640, 1598, 1090, 1070 cm⁻¹; ¹H NMR (360 MHz, DMSO- α) δ 8.48 (s, 1H, H-8), 8.16 (s, 1H, H-2), 7.34 (br s, 2H, NH₂), 6.57 (s, 1H, H-2'), 5.09 (t, α) (the second substantial s

= 5.5 Hz, 1H, OH), 4.28 (t, J = 5.1 Hz, 1H, H-4′), 3.55 (m, 2H, H-8′), 3.22 (dd, $J_{S',6'}$ = 9.2 Hz, $J_{S',6'}$ = 6.1 Hz, 1H, H-5′), 2.41 (m, 1H, H-6′exo), 2.25 (m, 1H, H-7′exo), 2.11 (m, 1H, H-7′endo), 1.90 (m, 1H, H-6′endo); 13 C NMR (90 MHz, DMSO- d_6) δ 157.0 (C, C-6), 153.9 (CH, C-2), 150.2 (C, C-4), 139.3 (CH, C-8), 120.0 (C, C-5), 93.0 (CH, C-2′), 86.4 (CH, C-4′), 72.3 (C, C-1′), 63.0 (CH₂, C-8′), 50.2 (CH, C-5′), 30.5 (CH₂, C-7′), 20.3 (CH₂, C-6′). HRMS (ESI+) calcd for [C₁₂H₁₄ClN₅O₂ + N₃]⁺ 318.0728, found 318.0726.

(1'S,2'R,4'S,5'R)-6-Chloro-9-(1'-chloro-4'-hydroxymethyl-3'-oxabicyclo [3.2.0]hept-2-yl)-9H-purine (48). To an ice-cooled solution of 42 (40 mg, 0.09 mmol) in THF (3 mL) was added Et₃N · 3HF (44 μ L, 0.27 mmol), and the resulting solution was stirred 20 h at room temperature. Evaporation of the solvent gave a residue which was purified by column chromatography (EtOAc-hexane 2:1) to afford 48 (28 mg, 0.09 mmol, 95% yield) as a colorless oil: $[\alpha]_D$ -23.6 (c 0.55, CHCl₃); IR (ATR) 3345, 3110, 2945, 1590, 1561, 1336, 1206 cm $^{-1}$; ¹H NMR (360 MHz, CDCl₃) δ 8.74 (s, 1H, H-2), 8.43 (s, 1H, H-8), 6.58 (s, 1H, H-2'), 4.36 (ddd, $J_{4',8'}$ = 4.5 Hz, $J_{4',8'}$ = 3.2 Hz, $J_{4',5'}$ = 3.2 Hz, 1H, H-4'), 3.97 (dd, J_{gem} = 12.1 Hz, $J_{8',4'}$ = 3.1 Hz, 1H, H-8'), 3.81 (dd, J_{gem} = 12.1 Hz, $J_{8',4'}$ = 4.5 Hz, 1H, H-8'), 3.39 (m, 1H, H-5'), 3.15 (br s, 1H, OH), 2.90 (m, 1H, H-7'endo), 2.65 (m, 2H, H-6'exo, H-7'exo), 1.99 (m, 1H, H-6'endo); 13 C NMR (90 MHz, CDCl₃) δ 151.9 (CH, C-2), 151.3/151.2 (2C, C-4/C-6), 144.0 (CH, C-8), 131.8 (C, C-5), 92.8 (CH, C-2'), 86.9 (CH, C-4'), 72.0 (C, C-1'), 63.4 (CH₂, C-8'), 50.2 (CH, C-5'), 34.1 (CH₂, C-7'), 21.4 (CH₂, C-6'). HRMS (ESI+) calcd for $[C_{12}H_{12}Cl_2N_4O_2 + H]^+$ 315.0410, found 315.0405.

(1'S,2'R,4'S,5'R)-9-(1'-Chloro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-adenine (52). A solution of 48 (15 mg, 0.05 mmol) and saturated NH₃/MeOH (4 mL) was heated at 90 °C in a sealed tube for 70 h. After cooling at room temperature, the solvent was removed under vacuum, and the resulting residue was purified by column chromatography (CH₂Cl₂-MeOH, 10:1) to afford 52 (12 mg, 0.04 mmol, 92% yield) as a white solid: mp 233–235 °C (from MeOH); $[\alpha]_D$ –36.0 (c 0.25, MeOH); IR (ATR) 3328, 3104, 2926, 1726, 1606, 1300, 1081, 1014 cm⁻¹; ¹H NMR (360 MHz, DMSO- d_6) δ 8.29 (s, 1H, H-8), 8.14 (s, 1H, H-2), 7.30 (br s, 2H, NH₂), 6.56 (s, 1H, H-2'), 5.13 (t, $J_{OH,8'} = 6.0$ Hz, $J_{OH,8'} =$ 5.7 Hz, 1H, OH), 4.29 (ddd, J = 4.4 Hz, J = 3.6 Hz, J = 3.3 Hz, 1H, 1H, 1H, 1H3.60 (m, 2H, H-8'), 3.24 (m, 1H, H-5'), 2.86 (m, 1H, H-7'), 2.45 (m, 2H, H-6'exo, H-7'), 1.99 (m, 1H, H-6'endo); ¹³C NMR (90 MHz, DMSO d_6) δ 156.9 (C, C-6), 153.7 (CH, C-2), 149.9 (C, C-4), 139.5 (CH, C-8), 119.4 (C, C-5), 91.6 (CH, C-2'), 86.6 (CH, C-4'), 73.5 (C, C-1'), 63.2 (CH₂, C-8'), 51.2 (CH, C-5'), 34.4 (CH₂, C-7'), 21.7 (CH₂, C-6'). HRMS (ESI+) calcd for $[C_{12}H_{14}CIN_5O_2 + H]^+$ 296.0909, found 296.0913.

(1'S,2'S,4'S,5'R)-9-(1'-Chloro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-hypoxanthine (55). To a solution of 47 (37 mg, 0.12 mmol) and MeONa (27 mg, 0.47 mmol) in dioxane (2 mL) were successively added 2-mercaptoethanol (33 µL, 0.47 mmol) and 2 drops of H₂O at room temperature. The solution was heated at 110 °C for 17 h when the same amount of MeONa, 2-mercaptoethanol, and H2O was added, and heating was continued for 2 days. Then, the solution was neutralized with glacial AcOH. The solvent was removed under vacuum, and the residue was purified by column chromatography (EtOAc-MeOH 9:1) to afford 55 (26 mg, 0.09 mmol, 74% yield) as a white solid: mp 206–208 °C (MeOH); $[\alpha]_D$ +35.0 (c 0.80, MeOH); IR (ATR) 3276, 3102, 3063, 2921, 2859, 1684, 1094 cm⁻¹; ¹H NMR (360 MHz, DMSO d_6) δ 12.43 (br s, 1H, N-H), 8.44 (s, 1H, H-8), 8.08 (s, 1H, H-2), 6.54 (s, 1H, H-2'), 5.08 (br s, 1H, OH), 4.29 (t, J = 4.8 Hz, 1H, H-4'), 3.54 (m, 2H, H-8'), 3.23 (dd, $J_{5',6'}$ = 8.9 Hz, $J_{5',6'}$ = 6.4 Hz, 1H, H-5'), 2.40 (m, 1H, H-6'exo), 2.25 (m, 1H, H-7'exo), 2.06 (m, 1H, H-7'endo), 1.88 (m, 1H, H-6' endo); 13 C NMR (90 MHz, DMSO- d_6) δ 157.5 (C, C-6), 148.9 (C, C-4), 147.2 (CH, C-2), 138.8 (CH, C-8), 125.1 (C, C-5), 93.4 (CH, C-2'), 86.7 (CH, C-4'), 72.4 (C, C-1'), 63.1 (CH₂, C-8'), 50.3 (CH, C-5'), 30.5 (CH₂, C-7'), 20.3 (CH₂, C-6'). HRMS (ESI+) calcd for [C₁₂H₁₃ClN₄O₃ + Na]⁺ 319.0568, found 319.0558.

(1'S,2'R,4'S,5'R)-9-(1'-chloro-4'-hydroxymethyl-3'-oxabicyclo[3.2.0] hept-2-yl)-9H-hypoxanthine (56). To a solution of 48 (10 mg, 0.03 mmol) and MeONa (7 mg, 0.12 mmol) in dioxane (2 mL) were successively added 2-mercaptoethanol (9 μ L, 0.12 mmol) and 2 drops of H₂O at room temperature. The solution was heated at 110 °C for 16 h and then, after being cooled to room temperature, was neutralized with glacial AcOH. The solvent was removed under vacuum, and the residue was purified by column chromatography (EtOAc-MeOH, 10:1) to give 56 (9 mg, 0.03 mmol, 96% yield) as a white solid: mp 130-133 °C (MeOH); $[\alpha]_D$ –18.0 (c 0.50, MeOH); IR (ATR) 3345, 3050, 2869, 1684, 1210 cm⁻¹; ¹H NMR (360 MHz, DMSO- d_6) δ 12.40 (br s, 1H, N-H), 8.25 (s, 1H, H-8), 8.07 (s, 1H, H-2), 6.53 (s, 1H, H-2'), 5.08 (br s, 1H, OH), 4.21 (ddd, $J_{4',8'} = 5.2$ Hz, $J_{4',8'} = 4.7$ Hz, $J_{4',5'} = 3.6$ Hz, 1H, H-4'), 3.61 (dd, $J_{gem} = 11.9 \text{ Hz}$, $J_{8',4'} = 4.7 \text{ Hz}$, 1H, H-8'), 3.56 (dd, $J_{gem} =$ 11.9 Hz, $J_{8',4'}$ = 5.2 Hz, 1H, H-8'), 3.23 (m, 1H, H-5'), 2.82 (m, 1H, H-7'exo), 2.45 (m, 2H, H-6'exo, H-7'exo), 1.98 (m, 1H, H-6'endo); ¹³C NMR (62.5 MHz, DMSO- d_6) δ 156.5 (C, C-6), 147.6 (C, C-4), 146.1 (CH, C-2), 137.9 (CH, C-8), 123.8 (C, C-5), 90.8 (CH, C-2'), 85.9 (CH, C-4'), 72.5 (C, C-1'), 62.1 (CH₂, C-8'), 50.1 (CH, C-5'), 33.4 (CH₂, C-7'), 20.5 (CH₂, C-6'). HRMS (ESI+) calcd for $[C_{12}H_{13}CIN_4O_3 +$ Na]⁺ 319.0568, found 319.0551.

ASSOCIATED CONTENT

Supporting Information. ¹H and ¹³C NMR spectra of all new compounds and 2D NMR spectra for compounds 32–33 and 49–56. This material is available free of charge via the Internet at http://pubs.acs.org.

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■ DEDICATION

Dedicated to the memory of Prof. Rafael Suau.

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